

Alma Mater Studiorum – Università di Bologna

DOTTORATO DI RICERCA IN

Oncologia e Patologia Sperimentale

Ciclo XXVI

Settore Concorsuale di afferenza: 06/A2

Settore Scientifico disciplinare: MED04

TITOLO TESI

**Genetic and environmental factors associated with
cardiovascular diseases and acute myocardial
infarction.**

Presentata da: Dott.ssa Manuela Ianni

Coordinatore Dottorato

Relatore

Prof. Sandro Grilli

Prof. Federico Licastro

Esame finale anno 2014

INDEX

-Introduction	pag. 4
Epidemiology.....	pag. 5
Classical risk factors.....	pag. 6
New risk factors.....	pag. 9
Inflammation.....	pag. 11
Genetic risk factors.....	pag. 14
Familiarity of CVD.....	pag. 17
Environmental factors.....	pag. 20
-Aim of the Study	pag. 24
-Materials and Methods	pag. 26
Subjects and patients.....	pag. 26
DNA extraction.....	pag. 27
SNP detection.....	pag. 27
Detection of EBV DNA.....	pag. 28
Detection of HHV-6 DNA.....	pag. 29
Statistical analysis.....	pag. 29
-Results	pag. 30
Genotype and allele frequency.....	pag. 30
Association between the triple genotype and cardiovascular risk.....	pag. 33
Body mass index (BMI) and blood lipid profile.....	pag. 34
Prevalence of CVE after 24 years of follow up.....	pag. 37
Detection of EBV and HHV-6 DNA.....	pag. 38

-Discussionpag. 44
Familiarity and AMI....., pag. 44
Genetic variations and AMI....., pag. 45
Virus infections and AMI..... pag. 47

-Conclusionpag. 50

-Referencespag. 51

INTRODUCTION

Cardiovascular Diseases (CVD) are the most frequent cause of morbidity and the leading cause of death in western societies and worldwide; furthermore, it remain today a great social and public sanity problem in Italy, due to the high number of people suffering these pathologies and the need for longterm care and rehabilitation trials (Braunwald E. 1997). CVD includes heart and blood vessel disease and their pathogenesis is related to atherosclerosis. Atherosclerosis is a condition that develops when a substance called plaque builds up in the walls of the arteries. This buildup narrows the arteries, making it harder for blood to flow through. If a blood clot forms, it can stop the blood flow and induce to heart attack or stroke.

The atherosclerosis leads to the development of atherosclerotic disease, the most frequent cardiovascular pathology, and its manifestations, in particular ischemic disease (acute myocardial infarction (AMI) and angina pectoris) and cerebral disease (ischemic ictus and hemorrhagic ictus).

A heart attack or AMI occurs when the blood flow to a part of the heart is blocked by a blood clot. If this clot cuts off the blood flow completely, the part of the heart muscle supplied by that artery begins to die. Usually, this is because one of the coronary arteries that supplies blood to the heart develops a blockage due to an unstable buildup of white blood cells, cholesterol and fat. The event is called "acute" when it is sudden and serious. The two main ways to determine if a person had AMI are electrocardiograms (ECGs) that trace the electrical signals in the heart and testing the blood for substances associated with damage to the heart muscle. Common blood tests are creatine kinase (CK-MB) and troponin. ECG testing is used to differentiate between two types of AMI based on the shape of the tracing. When the ST section of the tracing is higher than the baseline it is called an ST-elevation AMI (STEMI) which usually requires more aggressive treatment. Usually, when an event occurs in young age (35-45 years), the AMI is often fatal. In advanced age, often, the survivors to an AMI become a chronic patient that need continuous care all life long with high cost for society. Furthermore, the CVD are strongly associated to ageing and conduce to physically and cerebrally impairment.

An ischemic stroke happens when a blood vessel that feeds the brain gets blocked, usually, from a blood clot. When the blood supply to a part of the brain is shut off, brain cells will die. A hemorrhagic stroke occurs when a blood vessel within the brain bursts. The most likely cause is uncontrolled hypertension. The result will be the inability to carry out some of the previous functions as before like walking or talking. Some effects of stroke are permanent if too many brain cells die after a stroke due to lack of blood and oxygen to the brain. These cells are never replaced; others are only temporarily out of order.

Other types of CVD are: 1) Heart failure: means the heart is not pumping blood as well as it should, but the body's need for blood and oxygen is unchanged; 2) Arrhythmia: this is an abnormal rhythm of the heart that can beat too slow, too fast or irregularly, so the heart may not be able to pump enough blood to meet the body's needs; 3) Heart valve alterations: when heart valves do not open enough to allow the blood to flow through as it should two main alteration are observed valve stenosis and insufficiency.

Epidemiology

CVD kill an estimated 17 million people worldwide each year and over 4 million deaths in Europe. CVD weigh for 59% on global mortality versus 27% of tumor (World Health Organization. The top 10 causes of death, factsheet No. 310. Retrieved 14 August 2013 <http://who.int/mediacentre/factsheets/fs310/en/index.html>). CVD costs the Europe economy almost 196bn of euros per year (European Cardiovascular Disease Statistics 2012 Edition, p116. Retrieved 14 August 2013. <http://www.escardio.org/about/Documents/EU-cardiovascular-disease-statistics-2012>). In Italy, CVD are the most frequent cause of death and AMI is the main clinical complication of CVD.

In our country, the last data regarding the epidemiology of CVD come from *Istat* and *Istituto Superiore di Sanità* (ISS) in 2008. CVD cause 224.482 death per year (38,8% of total death). Of these, 33% are cardiovascular event (CVE) (AMI, ischemic heart diseases, angina pectoris and other chronic form of ischemic heart diseases).

Classical risk factors

The knowledge of the etiology and pathogenetic mechanisms of CVD is still limited and incomplete. Well-known traditional risk factors include associated with CVD are:

Age: the risk of developing CVD increases between 40-50 years for men and 50-60 for women;

Gender: the women are protected by sexual hormones up to the menopause and thereafter the percentage of CVD between women will increase. Therefore, gender is another important classical risk factor.

A family history of early heart disease: heart disease diagnosed before age 55 years in the father or a brother and before age 65 years in the mother or a sister increases the risk of CVD in offspring and sibling.

Race: African Americans are reported to be particularly at high risk to develop CVD than Asians, perhaps for the high levels of low-density lipoprotein cholesterol (LDL-C). The increased risk may be also associated with the different diets. Age, gender, race and familiarity are non-modifiable risk factors.

High cholesterol level: the Framingham Heart Study demonstrated that specially LDL-C, was the greater the risk of CVD. In 1984, the Lipid Research Clinics-Coronary Primary Prevention Trial revealed that lowering total and LDL (bad cholesterol levels) significantly reduced coronary heart disease (CAD). CAD was uncommon in people with cholesterol levels below 150 mg/dL. More recent series of clinical trials using statin drugs provided conclusive evidence that lowering LDL cholesterol reduces the rate of AMI, the need for percutaneous coronary intervention and the mortality associated with CAD-related causes (LaRosa JC et al., 2005)

Hypertension: in the Framingham Heart Study, even high-normal blood pressure (defined as a systolic blood pressure of 130-139 mm Hg, diastolic blood pressure of 85-89 mm Hg, or both) increased the risk of CVD 2-fold, as compared with healthy individuals (Vasan RS et al., 2001). A study by Allen et al. found that people who have increased or decreased in blood pressure during middle age had higher and lower remaining lifetime risk for CVD. This suggests that prevention

efforts should continue to emphasize the importance of lowering blood pressure in order to avoid hypertension (Allen N et al., 2012). Hypertension, along with other factors such as obesity contribute to the development of left ventricular hypertrophy (LVH). LVH founded to be an independent risk factor to CVD morbidity and mortality. It roughly doubles the risk of cardiovascular death in both men and women (Levy D et al., 1990).

Obesity: the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure (JNC VII) emphasizes weight control, adoption of the Dietary Approaches to Stop Hypertension (DASH) diet, with sodium restriction and increased intake of potassium and calcium-rich food, moderation of alcohol consumption to less than 2 drinks daily and increased physical activity (Chobanian AV et al., 2003). Obesity was associated with elevated vascular risk in population studies. In addition, this condition was associated with glucose intolerance, insulin resistance, hypertension, physical inactivity, and dyslipidemia (Rexrode KM et al., 1998). A study by Das et al examined more than 50,000 patients from the National Cardiovascular Data Registry with STEMI. The results suggested that although patients who were extremely obese (body mass index [BMI] >40) present at a younger age with STEMI, they have less extensive CAD and better LV function. However, as expected, their in-hospital mortality following STEMI was increased (adjusted odds ratio, 1.64) (Das SR et al., 2011).

Diabetes Mellitus: a disorder of metabolism associated with either insulin deficiency or insulin resistance. Glucose builds up in the blood stream, overflows through the kidneys into the urine, and results in the body losing its main source of energy, even though the blood contains large amounts of glucose. Patients with diabetes are 2-8 times more likely to experience future CVE than age-matched and ethnically matched individuals without diabetes (Howard BV et al., 2002). A recent study suggested a potential reduction of all-cause and CVD mortality in women with diabetes mellitus who consumed whole-grain and bran (He M et al., 2010). Another study suggested that meat consumption was associated with a higher incidence of coronary heart disease and diabetes mellitus (Micha R et al., 2010). A meta-analysis performed by Nordmann et al founded that the Mediterranean diet had more favorable changes in weighted mean differences

of body weight, body mass index, systolic blood pressure, diastolic blood pressure, fasting plasma glucose, total cholesterol, and high-sensitivity C-reactive protein than low-fat diets (Nordmann AJ et al., 2011).

Sigarettes smoking: Cessation of cigarette smoking constitutes the single most important preventive measure for CAD. As early as the 1950s, studies reported a strong association between cigarette smoke exposure and heart disease. Persons who consume more than 20 cigarettes daily have a 2- to 3-fold increase in total heart disease. Continued smoking is a major risk factor for recurrent heart attacks (Rea TD et al., 2002). Smoking is a risk factor for CVD in women and men; however, a systematic review and meta-analysis by Huxley and Woodward suggested that in some countries, smoking by women is on the rise; the study suggested that proper counseling and nicotine addiction programs should focus on young women (Huxley RR et al., 2011).

Physically exercise: The cardioprotective benefits of exercise include reducing adipose tissue, which decreases obesity; lowering blood pressure, lipids, and vascular inflammation; improving endothelial dysfunction, improving insulin sensitivity, and improving endogenous fibrinolysis (Thompson PD et al., 2003). In addition, regular exercise reduces myocardial oxygen demand and increases exercise capacity, translating into reduced coronary risk. Studies shown that even 15 minutes a day or 90 minutes a week of moderate-intensity exercise may be beneficial (Greenland P et al., 2010).

Modifiable risk factors are cigarettes smoke, diabetes, hypertension, hypercholesterolemia, high Body Mass Index (BMI), obesity, sedentary life-style changing the diet and follow a good life-style habit. Many sanitary preventative interventions of ISS to promote the good life style, to stop smoking, improve physically activities, and eat better (few salt, fat, or sweet) have decreased the incidence of CVD, in fact, in the last 40 years the mortality for CVD it's the half.

It was observed that more than half of patients with atherosclerotic complications, such as AMI, does not demonstrate classical risk factors.

New risk factors

Others factors are now considered as new risk factors (Ridker PM, 1999, Rifai et al, 2001).

C-reactive protein (CRP): is a protein in the blood that demonstrates the presence of inflammation, which is the body's response to injury or infection; CRP levels rise if inflammation is present. The inflammation process appears to contribute to the growth of arterial plaque, and in fact, inflammation characterizes all phases of atherothrombosis and is actively involved in plaque formation and rupture. According to some research results, high blood levels of CRP may be associated with an increased risk of developing CAD, CVE and having a heart attack (Arroyo-Espliguero R et al., 2004; Rifai N et al., 2001). In the Jupiter trial, in healthy persons without hyperlipidemia but with elevated high-sensitivity CRP levels, the statin drug rosuvastatin significantly reduced the incidence of major CVE (Ridker PM et al., 2008). The 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults states that measurement of CRP can be useful in selecting patients for statin therapy and may be reasonable for cardiovascular risk assessment, depending on the patient's age and risk level. CRP measurement is not recommended for cardiovascular risk assessment in asymptomatic high-risk adults, low-risk men 50 years or younger, or low-risk women 60 years or younger (Wang TJ et al., 2006).

An elevated lipoprotein(a) [Lp(a)] level is an independent risk factor of premature CAD (Hjemdahl P, 2002; Braunwald E, 1997) and is a significant risk factor for premature atherothrombosis and CVE. Measurement of Lp(a) is more useful for young individuals with a personal or family history of premature vascular disease and repeat coronary interventions. The 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults states that, in asymptomatic intermediate-risk adults, lipoprotein-associated phospholipase A2 might be reasonable for cardiovascular risk assessment (TJ et al., 2006). Lp(a) may be used to identify people at increased cardiovascular risk, but as of yet, there have been no studies on Lp(a) lowering because of the lack of available agents that are effective in reducing this value. Therefore, LDL lowering is probably the best strategy in people with elevated Lp(a) levels.

(Arrojo et al., 2004).

Homocysteine: a natural by-product of the dietary breakdown of protein methionine. In the general population, mild to moderate elevations are due to insufficient dietary intake of folic acid. Homocysteine levels may identify people at increased risk of heart disease, but again, due to the lack of agents that effectively alter the homocysteine levels, studies have not shown any benefit from lowering the homocysteine level (Miller, A et al., 1997). Levels of fibrinogen: an acute-phase reactant, increase during an inflammatory response. This soluble protein is involved in platelet aggregation and blood viscosity, and it mediates the final step in clot formation. Significant associations were found between fibrinogen level and risk of CVE in the Gothenburg, Northwick Park, and Framingham heart studies (Wilhelmsen L et al., 1984; Levenson, J., et al., 1995).

Low serum testosterone levels have a significant negative impact on patients with CAD. More studies are needed to assess better treatment (Malkin CJ et al., 2010). One meta-analysis suggested that the presence of erectile dysfunction increases the risk of CVD, coronary heart disease, stroke, and all-cause mortality. This additional risk may be independent of conventional cardiovascular risk factors (Dong JY et al., 2011).

One study suggested that women aged 50 years or younger who undergo a hysterectomy were at an increased risk for CVD later in life (Ingelsson E et al., 2011). Oophorectomy also increases the risk for both coronary heart disease and stroke.

A prospective cohort study (n=2312) by Kestenbaum et al evaluated older patients without CAD over 14 years. Vitamin D and parathyroid hormone were measured and the outcomes included AMI, heart failure, cardiovascular death, and all-cause mortality. Vitamin D deficiency was associated with increased mortality and AMI (each 10 ng/mL drop in vitamin D was associated with 9% greater increase in death and 25% increase in AMI). Parathyroid hormone excess was associated with a 30% increased risk of heart failure. Further randomized controlled trials are required (Kestenbaum B et al., 2011).

Inflammation

Inflammation appears to be important in the pathogenesis of atherosclerotic disease, since atherosclerotic plaques and lesions are associated with infiltration of activated immune cells and increased expression and synthesis of inflammatory markers (Hansson GK. 2001).

Cytokines are produced in a variety of tissues and regulate the expression of a number of inflammatory molecules, leading to destabilization and finally rupture of vulnerable atheromatic plaques. They also participate in the pathophysiology of acute coronary syndromes by direct effects on myocardial contractility and apoptosis. Several lines of evidence indicate that increased inflammatory cytokine levels and inflammatory cytokine activity detectable in peripheral blood have a prognostic role since they are useful markers predicting future CVE in patients with advanced atherosclerosis and in patients after coronary syndromes and may be useful in identifying subjects with a worse clinical course (Tousoulis D et al., 2006; De Gennaro L et al., 2012).

However, the existing data are limited and controversial. Potentially thousands of molecules are relevant to this apparently complex disease. Research paid much attention to sensitive specific serum biomarkers for vulnerable plaques as diagnostic tools for the identification of patients with acute coronary syndrome, but also to help us to identify high-risk patients. Furthermore, AMI is a myocardial necrosis occurring due to persistent coronary ischemia, in which inflammation plays an important role and heart failure is a common complication.

Elevated plasma levels of interleukine (IL-) 6 associated with increased risk of future AMI in a prospective study that involving 14916 apparently healthy men were found. These data thus support a role for cytokine-mediated inflammation in the early stages of atherogenesis (Ridker PM et al., 2000). IL-6 plasma levels was also associated with increased atherosclerosis when the control group was compared with the group free of subclinical CVD (Jenny, N.S et al., 2002).

IL-6 levels differentiated those with subclinical CVD from those without. Although the -174C allele was not associated with incident events, associations of the genotype with inflammation and AMI, combined with the plasma IL-6 results, suggest that IL-6 may chronically predispose an

individual to develop atherosclerosis.

Chiappelli et al. suggested that the C allele of the promoter polymorphism in the IL-6 gene is a risk factor for AMI in the elderly and the production of the IL-6 differentially affected different genotypes of the IL-6 -174 promoter polymorphism (Chiappelli M et al., 2005).

Elevated admission level of IL-6, but not of soluble CD40 ligand (sCD40L), metalloproteinase-9 (MMP-9), or tissue inhibitor of metalloproteinase-1 (TIMP-1), might indicate the onset of STEMI, and could provide prognostic value for future cardiac mortality within 2 y in patients with STEMI (Tan J et al., 2008).

A significant increase in IL-6 serum level as well as a significant decrease in IL-1 receptor alpha (Ra) for patients with a history of AMI was found. A trend toward higher concentration of inflammatory mediators was noticed in relation to the increase in severity of the aortic valve disease. An "inflammatory pattern" was associated with aortic sclerosis pathology and these results suggested the persistence of a chronic inflammation in patients who experienced acute coronary events (Rugina M et al., 2007).

The frequencies of genetic apo E (APOE) isoforms E2, E3 and E4 were determined in 523 patients with AMI. Significant difference in the frequency of APOE4 but not in the frequencies of isoforms E3 and E2 between patients and controls were observed (Utermann G et al., 1984). Several studies confirmed these results (Bennet AM et al., 2007).

Admission levels of serum MMP-9 and TIMP-1 closely correlated with left ventricular structure and function, which may be involved in the process of post-infarction remodeling of myocardium in AMI (Tan J et al., 2012).

Tumor Necrosis Factor alpha (TNF- α) may be an early marker of myocardial damage because of the early increase of its level after ischemic injury instead of being late consequence of extensive tissue necrosis. A significant increase was still seen at 48 hours post admission in patients with signs of heart failure but not in those without signs of heart failure. A significant positive correlation was found between plasma TNF-alpha level and calcium-dependent protein kinases level at admission. TNF- α level may be also an important indicator of the severity of AMI and the

occurrence of heart failure (Fahim MR et al., 2004). Plasma concentrations of TNF- α are persistently elevated among post-AMI patients at increased risk for recurrent coronary events. These data support the hypothesis that a persistent inflammatory instability is present among stable patients at increased vascular risk (Ridker PM et al., 2000a).

Monocyte chemoattractant protein-1 (MCP-1) appears to play a critical role at multiple stages in atherosclerosis, including the initiation of the fatty streak, promotion of plaque instability, and remodeling after myocardial infarction. In a large cohort of patients with acute coronary syndromes, an elevated baseline level of MCP-1 was associated both with traditional risk factors for atherosclerosis as well as an increased risk for death or AMI. For this reason, it appears to play a crucial role at multiple stages of atherosclerosis (De Lemos JA et al., 2003).

Between the pro- and anti-inflammatory components of the immune system there is a dynamic balance, but it is known that violation of which is an important mechanism of development of many pathological states, in this case, cardiac insufficiency. In the acute phase of AMI, inflammatory activation is enhanced with predominant proinflammatory response. In the course of the healing process within 6 months inflammation is suppressed and the balance between pro- and anti-inflammatory activation is restored. The size of AMI, BMI, lipid levels and the baseline levels of inflammatory markers influence the levels of inflammatory factors (Karpiński Ł et al., 2009).

Balance between the pro- and anti-inflammatory cytokines reflects the index of inflammatory activity. A study demonstrated that uncomplicated AMI patients, there is initially a high level of both proinflammatory and anti-inflammatory cytokines. After 10 days, were observed a decline in the IL-6 level and an increase in the TNF- α and IL-10 levels. In complicated AMI, as compared to uncomplicated AMI patients were observed initially high levels of TNF- α and IL-6 and reduced levels of IL-8 and IL-10, patients. In dynamics of supervision of this group, further increase of all proinflammatory cytokines and a decline of IL-10 was registered (Havrylenko TI et al., 2012).

Soluble interleukin (IL)-2 receptor (sIL-2r), IL-6, and IL-8 are increased in patients with acute

coronary syndrome and systolic dysfunction or acute heart failure (De Gennaro L et al., 2012). Some findings suggested that IL-8 and IL-12 was involved in the process of ischemic heart disease, and serum IL-12 may be a marker for differentiating AMI from unstable angina pectoris (Zhou RH et al., 2001). A number of studies associated blood levels of individual proinflammatory plasma biomarkers and gene variants that favor inflammation or cholesterol synthesis with AMI, but until now, none of these factors alone are sufficient to define risk for individuals. Furthermore, phenotype markers, blood levels of a given marker, may considerably vary with gender, age, concomitant diseases, metabolic disorders and diet, be sensitive to other environmental variables and significantly change with time in the same subject.

Genetic risk factors

Plasma levels of lipids and proinflammatory molecules fluctuate over time and in relation to age, sex, diet, lifestyle, medication, and environmental factors, limiting their usefulness in etiologic and predictive contexts. Inherited gene variants are, in contrast, constant and might provide a better indication of high intrinsic risk. Genetic markers may be used to overcome these limitations. For instance, gene polymorphisms (SNPs), known to influence the phenotypic expression of a given inflammatory molecule, may become informative in assessing individual risk of CVE and recent studies have focused on the genetic background of inflammatory cytokines as possible intrinsic risk factors associated with CVD. These investigations assessed gene variations largely distribute in human population and showed that some alleles of cytokine genes were more frequently distributed in patients with CVD than in controls (Humphries SE et al., 2001; Andreotti F et al., 2002).

Cytokines as regulators of activity of inflammation play significant role in mechanisms of formation of atherosclerotic plaques and in processes of their destabilization. One of leading genetic factors determining level of their production appears to be polymorphism of cytokine genes structure at their promoter loci (Konenkov VI et al., 2012).

Recent genome-wide association (GWA) studies have contributed substantially to the discovery

of new SNPs associated with CHD and AMI (Patel RS et al., 2011), but their clinical relevance is still unclear because a single gene variant can make a limited contribution to the total genetic load of AMI, and both common and rare gene polymorphisms may differentially affect susceptibility to the disease.

Data from a cross-sectional study suggested that the functional interaction of three genes the allele C of IL-1b, the allele C of IL-6 and e4 allele of APOE was strongly associated with AMI, affects pathogenetic mechanisms and an impaired regulation of immune responses plays a pivotal role in the disease (Licastro F et al., 2004). In the setting of non-ST-elevation acute coronary syndromes, genetic variation at the IL-1 gene locus contributes to the changes in soluble markers of endothelial inflammation (Ray KK et al., 2002).

A number of studies explored the association of ischaemic heart disease with SNPs of the inflammatory molecules TNF- α and β , transforming growth factors (TGF) beta1 and 2, IL-1 and its receptor antagonist (IL-1ra), the receptor for lipopolysaccharide (CD14), P and E selectins, and platelet endothelial cell adhesion molecule (PECAM) 1. The data provided some evidence that alterations in the genetics of the inflammatory system may modify the risk of ischaemic heart disease (Andreotti F et al., 2002).

Several gene variants that promote inflammation and cholesterol metabolism were associated with AMI. Promoter SNPs with functional relevance in the expression of the cognate inflammatory gene are often found at elevated frequency among patients with AMI. AMI across a broad age range' carried multiple proinflammatory alleles (IL6, TNF, IL10, Alpha 1-antichymotrypsin (ACT)). 'A subset of AMI in middle age' had numerous proinflammatory alleles (IL6, TNF, ACT, interferon gamma IFN- γ , 3-hydroxy-3-methyl-glutaryl-CoA reductase (HMGCR)). 'AMI after age 80' had a reduced risk set (IL6, IL10, IFN- γ) (Licastro F et al., 2011; Licastro F et al., 2007).

IL-10 reflects a proinflammatory state in patients with acute coronary syndrome and was suggested that IL-10 is as effective a biomarker for the risk prediction of future CVE as other markers of systemic inflammation (Målarstig A et al., 2008). Also IL-8 -251 A/T polymorphism is

associated with acute coronary syndrome risk in the Chinese Han population and the A allele of IL-8 -251 A/T may be an independent predictive factor for acute coronary syndrome (Zhang X et al., 2011). IL-18 gene is a susceptibility locus for AMI, a finding of potential interest in the clinical practice (Koch W et al., 2011a).

Six different and functionally relevant SNPs of the genes coding for IL-10, TNF- α , and TNF- β are neither separately nor in cooperation associated with the risk of CAD or AMI in angiographically examined study patients (Koch W et al., 2001b). Also a relationship among genetically determined TNF- α and pro-thrombin (FII) production and increased levels of tissue damage markers of AMI, suggest that a complex genetic background, might be involved in susceptibility to AMI in young men influencing the extension and severity of the disease (Vaccarino L et al., 2013).

Although many risk factors are intercorrelated, raising the possibility of a higher level of genetic control by a small number of master genes that control fundamental physiological systems. Such genes are likely to be relevant to the combined processes of plaque instability and coronary thrombosis.

The complex pathogenesis of AMI implicates phenotypic and genetic heterogeneity. A new mathematical algorithm named Auto Contractive Map (AutoCM), was applied in AMI patients to detect and evaluate the relationships among genetic factors, clinical variables and classical risk factors. Genes were selected because their strong regulatory effect on inflammation and SNP in these gene were located in the promoter region. In the connectivity map generated by AutoCM a group of variables was directly linked with the AMI status: gender (male), early age at onset (50-65 years), HMGCR gene (CC wild type genotype), IL-1 β , ACT, IL-6 GG and vascular endothelial growth factor (VEGF) CC genotypes. This direct link suggested a possible pathogenetic association with AMI. Other genetic, clinical and phenotypic variables were associated to the disease under a statistically defined hierarchy showed in the new connectivity map generated by AutoCM. These analyses suggested also that genotypes of few inflammatory genes, a SNP in HMGCR gene, middle age, gender, low HDL and diabetes were very informative variables to

predict the risk of AMI (Licastro F et al., 2010).

Familiarity of CVD

Parental history is a widely accepted risk factor for offspring CVE, although the mechanisms remain unclear. Many studies examined the contribution of conventional and genetic risk factors in explaining the excess risk of CVE in offspring with positive parental history. Whether parental CVD confers increased risk independent of other risk factors remains controversial. Prior studies relied on offspring report, without complete validation of parental events.

Despite some study underline an association of positive parental history with blood pressure, total and high-density lipoprotein cholesterol, CRP, and physical activity, less than 15% of the risk was explained through conventional and novel risk factors. The greatest risk of CVD was observed in positive parental history participants with elevated CRP or hypertension and substantially increases the risk of CVD (Hamer M et al., 2009).

Genotypes of inflammatory molecules seem to play an opposite role in atherosclerosis and longevity: pro-inflammatory genotype seems to contribute significantly to the risk of coronary heart disease, but some study supported the hypothesis that the genetic background favoring CVD. So several study in centenarian verify the role of proinflammatory alleles, such as pyrin and C-C chemokine receptor type 5 (CCR5), in AMI and longevity. The results suggested the genetic background favoring CVD and that the centenarian genetic background may be useful for investigating genetic key components of age-associated diseases as atherosclerosis (Candore G, et al., 2006).

A family history of CVD and AMI is frequently encountered in clinical practice, and a premature heart attack in parents is associated with a high risk of the disease in their offspring. A small subgroup of children, whose parents suffered a heart attack in their late thirties and early forties, may be at particularly high cardiovascular risk was evaluated about weight, blood pressure, plasma cholesterol levels, obesity and smoking and it is concluded that cardiovascular risk factor screening among children with a positive family history of premature atherosclerotic

complications is appropriate and cost-effective (Rumboldt M et al., 2003).

To assess the risk of CVE, the diagnostic effectiveness of molecular-genetic markers (SNPs in APOE, ACE (I/D) and MTHFR (C677T) genes) in combination with conventional risk factors was studied. The combination of results of molecular-genetic testing with conventional risk factors allows to increase the predictability of cardiovascular risks (Nazarenko GI et al., 2009).

Some study examined the association of parental CVD with 8-year risk of offspring CVD, using pooled logistic regression. Parental CVD independently can predict future offspring events in middle-aged adults. Addition of parental information may help clinicians and patients with primary prevention of CVD, when treatment decisions may be difficult in patients at intermediate risk based on levels of single or multiple risk factors (Lloyd-Jones DM et al., 2004).

The relevance of familial factors to CVD is further supported by the finding that siblings with CVD are at increased risk of future CVE in middle-aged adults regardless of the presence of established risk factors (Murabito JM et al., 2005).

The associations of parental longevity with carotid intima-media thickness, carotid plaques, and aortic arterial stiffness in adult offspring was also examined and the results indicate that there are modifications of structure and function of large arteries according to paternal longevity (Zureik M et al., 2006).

Data from the longitudinal Framingham Heart Study confirmed that subjects with long-lived parents have a better cardiovascular risk profile in middle age than those whose parents died younger and centenarians have better cardiovascular risk profiles compared to younger old people (Terry DF et al., 2007).

Some reports revealed that CVD (i.e. hypertension, diabetes, angina and/or myocardial infarction) are less common in centenarians respect to 70 and 80 years old persons. A combination of genetic factors and lifestyle aspects appears to contribute to the longevity of centenarians. A role on this better cardiovascular risk profile may be played by the increasing use of pharmacologic treatments in the elderly population (especially for hypertension and dyslipidemia), but the contribution of drug treatments to promote extreme longevity is not confirmed. In fact,

centenarians in general needed fewer drugs at younger ages due to a healthy lifestyle. The importance of the genetic contribution was demonstrated by the inheritance of low-risk cardiovascular profiles in centenarian offspring and lower prevalence of cardiovascular CVD in this population as compared with their spouses or with age-matched subjects without centenarian parents (Galioto A et al., 2008).

Furthermore, the Offs of centenarians have a better cardiovascular risk profile and retain some important cardiovascular advantages over time than those of parents not enjoying a long life (Adams ER et al., 2008). It is interesting to note that structural vascular changes can be observed in Offs with a parental history of AMI at a young age, regardless of the presence of a number of the classical cardiovascular risk factors. Increased carotid intima-media thickness is an early manifestation of atherosclerosis. A parental history of premature AMI was associated with an increase in carotid intima-media thickness in children-adolescents and young adults. Offspring of coronary patients showed an unfavourable lipid profile compared to controls; however, the association between a premature AMI and carotid intima-media thickness was independent of lipids, apolipoproteins and other traditional risk factors. So, vascular structural changes are detectable in Offs of parents with premature AMI at a young age and occur independently of several traditional cardiovascular risk factors (Barra S et al., 2009).

Once again, the Framingham Heart Study shown that parental stroke before the age of 65 years is associated with a 3-fold increase in the risk of stroke in their Offs, and that parental history can be used as clinical risk marker of an individual's propensity to stroke (Seshadri S et al., 2010).

The distribution of some SNPs influencing a inflammation and found an higher prevalence of pro-inflammatory SNPs (SNP A2080G of pyrin gene, SNP Gly670Arg of PECAM gene, C1019T of Cx 37 gene, SNP G1059C of PCR gene) and a lower prevalence of anti-inflammatory SNPs (Asp299Gly of TLR4 gene, SNP -1082 G/A of IL10 gene, CCR5 Δ 32) in young patients with AMI was found. Results of these studies shown that early AMI could be associated with a genetic predisposition to an intense inflammatory response, also to an hyperviscosity syndrome (Incalcaterra E et al., 2010).

All these findings suggest that genetic studies of offspring may be useful to identifying transmissible genetic traits in CVD.

Environmental factors

Studies about associations of infections with viruses and other pathogens, with CVD, frailty and/or mortality are conflicting. Since high levels of antibodies against these pathogens occur in the elderly, the role of pathogens in morbidity and mortality of vulnerable elderly was explored. Increasing evidence supports a link between serological evidence of prior exposure to infectious pathogens, pathogen burden, and the risk for future AMI and death in patients with coronary artery disease. The role of viral infections in the pathogenesis of atherosclerosis remains controversial largely due to inconsistent detection of the virus in atherosclerotic lesions, but the identification of specific viral signatures can help to understand pathogenetic mechanisms in AMI.

In the past, evidence for enterovirus (EV), adenovirus, and cytomegalovirus (CMV) was in the focus of attention; in the meantime, "new" cardiotropic pathogens such as parvovirus B19, Epstein-Barr virus (EBV), and human herpesvirus (HHV-) 6 in patients with dilated cardiomyopathy with and without inflammation have been detected. Their persistence in the myocardium correlates with a decline in pumping capability within 6 months. While the virus is still being eliminated, the second phase of the disease begins, which is characterized by autoimmune phenomena and often a cardiac inflammatory response which likewise correlates with a worsening prognosis. The transition to the third and final phase with development of dilated cardiomyopathy occurs gradually and can take years. The goal of every diagnostic and therapeutic intervention must be to eradicate the virus and eliminate the inflammatory response to prevent the disease from progressing to terminal cardiac insufficiency (Pankuweit S et al., 2010). Some study evaluated the intimal presence of four pathogens in human coronary atheroma, clinically associated with acute coronary syndromes and stable angina and the effect of pathogen burden on the expression of human heatshock protein 60 (hHSP60), a key protein in

(auto-)immune pathogenesis of atherosclerosis. The data demonstrate that there exist an impact of intimal pathogen burden in plaque instability, and suggest the presence of (auto-)immunoreactions against upregulated hHSP60 is an important pathomechanism that may contribute to acute coronary syndromes (Andrié R et al., 2003).

A study assess the detection of human HHV-6 and HHV-8, two DNA viruses with a distinct tropism for endothelium and lymphocytes, with the aim to know if these virus may be associated with coronary instability and to found a correlation between infection-driven inflammatory burden and acute manifestation of coronary artery disease.

HHV-8 viremia was undetectable in all three groups of study: patients with AMI, patients with stable coronary artery disease and patients without coronary and carotid artery atherosclerosis subjected to cardiac valve replacement. HHV-6 viremia was detected in a substantial fraction of the samples examined without differences between groups (Magnoni M et al., 2012).

To evaluate the association of ischemic heart disease with the number of pathogens among individuals with infection serum samples of patients with ischemic heart disease as the AMI or unstable angina and healthy subjects for the presence of antibodies to *Helicobacter pylori* (*H. pylori*), CMV, HSV-1 and HSV-2 were tested. The rate of subjects with high infection burden (3 seropositivities) was significantly higher in ischemic heart disease group than to control group. The seroprevalence of anti-*H. pylori*, anti-CMV and anti-HSV-1 antibodies in AMI and unstable angina groups was significantly higher than control group. The infection burden was significantly increased in patients with ischemic heart disease and this parameter should be considered as an independent risk factor for development of ischemic heart disease (Jafarzadeh A et al., 2011).

Some study described also a dual role of infections as risk factors for coronary heart disease and showed that EV, HSV and *Chlamydia pneumoniae* (*C. pneumoniae*) HSP60 IgG antibody titers were associated with increased risk of coronary heart disease and protection from infections, usually suffered during the childhood before the era of MMR vaccination, may predispose the individual to coronary heart disease (Pesonen E et al., 2007).

The levels of IgG antibody specific for HSV-1, CMV and EBV among patients with

atherosclerotic vascular diseases were assessed to elucidate whether infectious agents such as herpes viruses could be implicated in the inflammatory atherosclerotic process and to examine the relation between the levels of these antibodies lipid profile and high-sensitive C-reactive protein (hsCRP) in these patients. The level of CMV- and EBV- specific antibodies were elevated among vascular disease patients and the presence of CMV-specific IgG was associated with the development of the disease. Serum lipids and hsCRP were increased among the patients; however, no significant correlation was detected between antiviral IgG levels and lipid profile or hsCRP (Al-Ghamdi A, 2012).

The *C. pneumoniae*, HSV-1, CMV and EBV DNA in atherosclerotic plaques from carotid arteries obtained from 17 patients were investigated. Genomic sequences of *C. pneumoniae*, HSV-1, CMV were not found in any atherosclerotic lesion. Therefore, these results did not support the hypothesis of an association between these infectious agents and atherosclerosis. Conversely, three patients were found to be positive for EBV DNA, thus indicating that, at least in a limited number of patients, EBV could play a role in atherogenesis (Tremolada S et al., 2011).

The intimal presence of 4 pathogens in atheroma, clinically associated with acute coronary syndromes and stable angina and the effect on the expression of intimal CRP, tissue factor and hHSP60 to demonstrate the impact of pathogen burden in plaque instability and suggest local proinflammatory, prothrombotic, and proimmunogenic effects was investigated. Analysis revealed *C. pneumoniae* was present in 73%, *H. pylori* in 31%, CMV in 16%, and EBV in 40%. Also the expressions of CRP, tissue factor, and hHSP60 were significantly higher in acute coronary syndromes lesions and the number of infectious pathogens correlated significant with the expressions of CRP, tissue factor and hHSP60 (Andrié RP et al., 2010).

In a cross-sectional study, the presence of *C. pneumoniae*, *Mycoplasma pneumoniae* (*M. pneumoniae*), CMV and EBV in atherosclerotic and non-atherosclerotic vascular samples was investigated. The results suggest that *C. pneumoniae*, *M. pneumoniae* and CMV were present with similar frequency both in atherosclerotic and non-atherosclerotic vessels (Bayram A et al., 2011).

The presence of HHV-6, HHV-7 and HHV-8 DNA in carotid, iliac and coronary artery specimens obtained from a group of subjects with atherosclerosis and a control group were investigated. HHV-6 but not HHV-7 and HHV-8 DNA were found in atherosclerotic vascular tissues. However, further research on broader study groups and with different protocols is needed to determine whether these viruses play a role in the formation of atherosclerosis (Kaklikkaya I et al., 2010).

Explanted heart of 28 patients, who underwent heart transplantation were screened for EV, Adenovirus Type 3 (AdV3) and HHV6 sequences in order to assess the incidence of cardiac viral infection that may be implicated in the pathogenesis of cardiomyopathy, and estimate viral distribution in the myocardium. AdV3 and HHV6 sequences coexisted in one case with inflammatory cardiomyopathy (Tátrai E et al., 2011).

The presence of CMV immediate early genomic region of DNA in 40 arterial specimens from coronary plaques and 27 samples from normal vessels were evaluated. CMV DNA was detected in 9 out of 26 patients (34.6%) in both non-atherosclerotic tissues and atherosclerotic plaques. No statistically significant differences were observed between normal and diseased vessels. These findings do not support a direct causative role of CMV in the development of atherosclerotic plaques (Xenaki E et al., 2009).

Some publications shed light to the role of AdV3, HHV- 6 and EV viruses in the development of dilated cardiomyopathy. AdV3, HHV- 6 and EV genomes in DNA and RNA were isolated from five regions of the heart muscle. In 2 patients AdV3 and in 1 patient both AdV3 and HHV-6 were detected. The AdV3 genome was the most frequent virus genome in explanted heart tissues. The identified viral sequences suggested previous viral infection, which could have played a role in the development of dilated cardiomyopathy (Tátrai E et al., 2007).

AIM OF STUDY

CVD are rising as a cause of death and disability worldwide and also a great social and medical problem and AMI is the main clinical complication. The knowledge about the aetiology and pathogenetic mechanisms of coronary vessels disease is limited and incomplete. AMI is a multifactorial disease with a complex pathogenesis in which lifestyles, individual genetic backgrounds and environmental risk factors contribute to the pathogenetic mechanisms and clinical manifestations.

More than half of patients with atherosclerosis do not show classical risk factors. New risk factors for CVD are emerging as the results of a growing understanding of the process of atherogenesis. Inflammation appears to be important in the pathogenesis of atherosclerotic disease, since atherosclerotic plaques and lesions are associated with infiltration of activated immune cells and increased expression and synthesis of inflammatory markers. Elevated blood levels of cytokines and interleukins in patients with CVD have been also found. However, phenotype markers, may considerably vary in several physiological condition or be sensitive to other environmental variables and significantly change with time in the same subject. Genetic markers may be used to overcome these limitations. For instance, gene polymorphism, known to influence the phenotypic expression of a given inflammatory molecule, may become informative in assessing individual risk of CVE.

Furthermore, it is known that a family history and a still largely undefined genetic background greatly influence the early clinical manifestation of AMI and CVD.

We investigated genetic variations represented by SNPs in the promoter region of a number of genes regulating metabolic and immune functions that have previously been found to be associated with an increased risk of AMI in case/control studies as possible genetic markers of an increased risk of CVD in children of parents with a positive history of AMI (Licastro F et al., 2010).

Three populations were studied: a population affect of AMI, unaffected subjects with positive

familiarity for AMI and a control group of patients without history of CVD to improve our understanding about genetic background involved in the development of atherosclerosis and its complications.

A second control group of comparable age from the WHO-MONICA Brianza study to compare CVE prevalence during the 24 years follow-up of our Offs was also used.

In second instance we investigated the same SNPs in genes in another group of AMI to confirm the obtained results.

Finally, DNA samples from peripheral blood leukocytes (PBL) of all patients were analyzed for the presence of EBV and HHV-6 to assess the involving of a viral etiology as risk factors of CVD and AMI.

MATERIALS AND METHODS

Subjects and patients

The study involved 154 Offs from Northern Italy, each of whom had one parent who had experienced an AMI before the age of 65 years. An evaluation was made of the classic AMI risk factors (metabolic parameters, smoking, diabetes, obesity, a sedentary lifestyle) together with ECG records, anthropometric indices, arterial pressure and medical history. The Offs were re-screened after a follow-up of 24 years in order to assess any changes in behavioral and biological risk factors and collect blood samples for laboratory and genetic evaluation. The CVD risk factors and drug treatments recorded at baseline and 24 years later were compared with those of the age-matched population of the WHO-MONICA Project, which were collected in Brianza, Northern Italy, during independent surveys carried out in 1984, 1991 and 2004 (Ferrario M et al., 2001).

A further group of 269 consecutive patients with a clinical diagnosis of AMI based on electrocardiographic changes and standard laboratory findings, and confirmed by echocardiography and coronary angiography (Licastro F et al., 2010), who were admitted to the Cardiology Unit of Ferrara University Hospital during 2006–2007, was investigated.

Another group of patients with cardiomyopathy (CMP) belonged from Cardiology Unit of Ferrara Hospital and included 177 AMI (patients that at the moment of hospitalization had enzymatic dismission index typical of myocardial necrosis) with two types of myocardial infarctions based on the shape of the tracing. An ST section of the tracing higher than the baseline is called an ST elevation AMI (STEMI) which usually requires more aggressive treatment and another one without ST elevation (NSTEMI: Non ST Elevation Myocardial Infarction) (Moe KT et al., 2010).

The healthy CTR were 315 subjects without a family history of CVD participating in the “Conselice study of brain aging” conducted in Northern Italy in 1999–2005 (Ravaglia G et al., 2001), none of whom showed any signs of CVD or inflammatory diseases before or during the study. The plasma cholesterol and lipid profiles of the Offs, AMI patients and CTR were determined on the basis of standard laboratory procedures. The research protocol was approved by our Institutional Review Boards, and all of the participants gave their written informed

consent.

DNA extraction

5 ml of blood diluted in a Falcon with $\frac{3}{4}$ volumes of Phosphate Buffer Saline 1X were extracted from the patients and were centrifuged for 20 minutes at a temperature of 4 ° C at 4000 rpm . 20 ml of saline solution Nonidet P40/NaCl (Nonidet P40 0,1% e NaCl 0,9%) were added to the pellet and it is centrifuged again for 15 minutes at 4 ° C at 4000 rpm . 5 ml of lyses buffer (Urea 7 M; NaCl 0,3 M; EDTA 10 mM; Tris-HCl 10 mM a pH 7,5) and then 1 ml of 10 % SDS were added to the pellet and incubated for the lysis in thermostat bath at 37 ° C for 10 minutes. 7 ml of phenol / chloroform / isoamyl alcohol 25:24:1 were added and centrifuged for 15 minutes at 4 ° C to 3900 rpm. the supernatant was collected it was measured the volume. 50 ml Sodium acetate is added to the supernatant to obtain a final concentration of 0.2 M. Then 2-3 volumes of 95% ethanol were added to the solution; shaking slightly, you can see a suspension of filamentous DNA (jellyfish). The suspension was centrifuged at 4 ° C at 3500 rpm for 20 minutes. To collect the DNA, the pellet coated into the tube was resuspended in 1 ml of 70% ethanol. The pellet thus collected was centrifuged at 13000 rpm for 10 minutes. Removed the ethanol, the pellet was dried and finally was resuspended in water by controlling the viscosity to add a quantity of water adequate. The concentration of DNA was read in a spectrophotometer at 260 nm . The samples obtained are maintained at -20 ° C (Grimaldi LM et al., 2000).

SNP detection

The presence of SNPs in the promoter regions of the VEGF (-2578 C/A, rs699947), ACT (-51 G/T, rs1884082), HMG-CR genes (-911 C/A, rs3761740), IL-1 β (-511 C/T, rs16944) and IL-10 (-1082 G/A, rs1800896) were detected by real-time PCR. The SNP-specific primers and probes were designed using the TaqMan genotyping assay (ABI, Foster City, CA) in a 25 μ l total volume of BIORAD CFX 96 in accordance with the manufacturer's instructions (Licastro F et al., 2010). IFN- γ genes (+874 T/A, rs2430561) genotype was assayed by RT-PCR using allele specific

modified LNA primers (TTTATTCTTACAACACAAAATCAAATC+T, TTTTATTCTTACAACACAAAATCAAATC+A, TGTGCCTTCCTGTAGGGTATTATTA). The polymorphic SNP was located at the 3'-position of the forward primers and a single-LNA base was incorporated at this position (+) (Proligo, Italy).

RT-PCR was performed in 96 well plates using a BIORAD CFX 96 platform. Reaction volume (25 µl), included a SYBR Green PCR Master Mix with the enzyme, Mg²⁺ and dNTP (ABI, Foster City, CA, USA; 200 nmol/L) PCR primers and genomic DNA (0.5 ng/µl). A start of 10 min at 95°C was followed by 40 cycles at 95°C for 15 s and 60°C for 60 s. (Licastro F et al., 2010).

Detection of EBV DNA

DNA samples were analyzed by qPCR. For the standard curve, we performed a nested PCR with an EBV-positive sample under the same conditions as described above. The PCR products were loaded onto a 2% agarose gel and purified with a QIA quick gel extraction kit (Qiagen). Concentration of DNA was determined using a Beckman DU 460 spectrophotometer in which the purified PCR product was placed onto the apparatus and the optical density (OD) measured ($\lambda = 260$ nm). OD values were converted to the appropriate concentrations (ng/µL). The following equation was used to calculate the copy numbers from a known PCR product concentration: $\text{weight of PCR fragment (g/}\mu\text{L)} / (660 \text{ g per mol} \times \text{number of base pairs of the PCR fragment}) \times (6.023 \times 10^{23}) = \text{number of genomic copies/}\mu\text{L}$ (Malorny B et al., 2003). Tenfold dilutions were made of this cleaned-up DNA. In each run we added virus-specific standards (10², 10³, 10⁴, 10⁵ copies/5 µL), which were used to generate the reference curve to quantify viral DNA in individual samples. Primer sequences and PCR cycling conditions are listed: GCCAGAGGTAAGTGGACTTTAAT, GAGGGGACCCTGAGACGGGT

(Amplicon size/protein: 96 bp), (PCR cycle conditions: 50 °C: 2 min, 95 °C: 15 min, 45 cycles of 94 °C: 2 min, 60 °C: 30 s, 72 °C: 30 s, melt curve 65 °C to 95 °C, 4 °C.), (Limited optical Detection: 33 copies/reaction).

Detection of HHV-6 DNA

DNA samples from peripheral leukocytes were analyzed by qPCR. Standard curve for HHV-6 was prepared as described above for the EBV standard curve. For the external PCR, 250 ng/5 μ L of DNA template was added to a 50- μ L mixture containing 1 μ mol/L of each external primer, 200 μ mol/L of dNTP (Fermentas), reaction Buffer 1X (Euroclone), MgCl₂ 2 mmol/L (Euroclone), and 2 U of *Thermus aquaticus* polymerase (EuroTaq, Euroclone). For the internal PCR, 10 μ L of the previous PCR reaction was added to a 50- μ L mixture under the same conditions as the external PCR. Primer sequences and PCR cycling conditions are listed: TCCATTATTTTGGCCGCATTCGT, TGTTAGGATATACCGATGTGCGT (Amplicon size/protein: 173 bp), (PCR cycle conditions and Limited optical Detection as described above for EBV).

Statistical analysis

The different genotypes were statistically analyzed using contingency tables and the chi-square (χ^2) test, and the odds ratios (OR) and confidence intervals (CI) and their statistical significance were also calculated. The level of statistical significance was set at 5%. The mean values of the various quantitative variables were compared by means of one-way analysis of variance (ANOVA) followed by appropriate posthoc comparisons and Bonferroni's correction. Statistical tests were two-sided, and significance was set at $p < 0.05$.

The Hardy-Weinberg equilibrium was verified for the two control groups. A logistic regression model was used to evaluate the effect of several clinical variables and the SNPs on the risk of CVD and AMI.

RESULTS

Genotype and allele frequency

Table 1 shows the SNP numbers, gene positions and mutated alleles of the investigated VEGF, ACT, HMGCR, IL-1 β , IL-10 and IFN- γ genes, along with the number, mean age and gender of the subjects in the different groups.

Table 1. SNPs list and the number, age and gender distribution of CTR, Offs, AMI and CMP

Gene polymorphism:			
VEGF	(rs 699947)	SNP at -2578, allele mutation = A	
IL-10	(rs 1800896)	SNP at -1082, allele mutation = A	
IFN- γ	(rs 2430561)	SNP at +874, allele mutation = A	
ACT	(rs 1884082)	SNP at -51, allele mutation = T	
HMG-CR	(rs 3761740)	SNP at -911, allele mutation = A	
IL-1 β	(rs 16944)	SNP at -511, allele mutation = T	
	N	Mean Age	Gender
CTR	321	72 \pm 5.1	158 M/ 163 F
Offs	154	55.8 \pm 6.7	80 M/ 74 F
AMI	267	67.7 \pm 12.2	195 M/ 72 F
CMP	396	70 \pm 11	347 M/90 F

The genotype distribution and allele frequency of the VEGF gene between Offs, AMI and CTR groups are shown in Table 2.

Table 2 Genotype distribution and allele frequency of VEGF SNP (rs 699947) from Offs, CTR and AMI

VEGF	CC (n) %	CA (n) %	AA (n) %	C carriers (n) %	A carriers (n) %
Offs (n = 154)	(97) 63	(42) 27.3	(15) 9.7	(139) 90.3	(58) 37.7
CTR (n = 291)	(119) 40.9	(127) 43.6	(45) 15.5	(246) 84.5	(172) 59.1
AMI (n = 257)	(167) 65	(78) 30.4	(12) 4.7	(245) 95.3	(90) 35
Offs vs CTR	$\chi^2 = 19.680$, $p = 0.0001$; CC carriers vs non-CC carriers: $\chi^2 = 19.745$, $p = 0.0001$; A carriers $\chi^2 = 18.545$, $p = 0.0001$.				
AMI vs CTR	$\chi^2 = 36.906$, $p = 0.0001$; CC carriers vs non-CC carriers: $\chi^2 = 31.796$, $p = 0.0001$; OR = 2.689 (CI: 1.900-3.806); AA carriers vs non-AA carriers: $\chi^2 = 16.943$, $p = 0.0001$; OR = 0.269 (CI: 0.139-0.521); C carriers $\chi^2 = 17.063$, $p = 0.0001$; OR = 3.735 (CI: 1.929-7.232); A carriers $\chi^2 = 31.733$, $p = 0.0001$; OR = 0.373 (CI: 0.264-0.527).				
Offs vs AMI	$\chi^2 = 4.141$, $p = 0.126$.				

The CC genotype was more frequent in the Offs than in CTR (63% vs 40.9%, $p = 0.0001$), and also more frequent in the AMI group (65% vs 40.9%, $p = 0.0001$; OR = 2.689). The percentage of VEGF C carriers was significantly higher in the AMI group than in the CTR (95.3% vs 84.5%, $p = 0.0001$; OR = 3.735), whereas the percentage of A carriers was significantly lower in the Offs (37.7%, $p = 0.0001$) and the AMI group (35%, $p = 0.0001$; OR = 0.373) than the CTR (59.1%). The AA genotype was more frequent in the CTR than in the AMI group (15.5% vs 4.7%, $p = 0.0001$; OR = 0.269).

The genotype distribution and allele frequency of the IL-10 gene between Offs, AMI and CTR groups are shown in Table 3.

Table 3 IL-10 SNP (rs 1800896) genotype distribution and allele frequency from Offs, CTR and AMI

IL-10	GG (n) %	GA (n) %	AA (n) %	G carriers (n) %	A carriers (n) %
Offs (n = 154)	(20) 13	(80) 51.9	(54) 35.1	(101) 65.6	(135) 87.7
CTR (n = 239)	(73) 30.5	(88) 36.8	(78) 32.6	(161) 67.4	(165) 69
AMI (n = 265)	(56) 21.1	(141) 53.2	(68) 25.7	(197) 74.3	(209) 78.9
Offs vs CTR	$\chi^2 = 17.378$, $p = 0.0001$; GG carriers vs non-GG carriers: $\chi^2 = 15.981$, $p = 0.0001$; A carriers $\chi^2 = 17.984$, $p = 0.0001$				
AMI vs CTR	$\chi^2 = 13.887$, $p = 0.001$; GG carriers vs non-GG carriers: $\chi^2 = 5.845$, $p = 0.016$; OR = 0.609 (CI: 0.407-0.912); A carriers $\chi^2 = 6.344$, $p = 0.012$; OR = 1.674 (CI: 1.179-2.504)				
Offs vs AMI	$\chi^2 = 6.550$, $p = 0.038$; GG carriers vs non-GG carriers: $\chi^2 = 4.352$, $p = 0.037$; OR = 1.795 (CI: 1.031-3.126); AA carriers vs non-AA carriers: $\chi^2 = 4.077$, $p = 0.043$; OR = 0.642 (CI: 0.418-0.989); A carriers $\chi^2 = 5.126$, $p = 0.024$; OR = 0.525 (CI: 0.299-0.923).				

Table 3 shows the IL-10 genotype distribution and allele frequency in the three groups. The GG genotype was more frequent in the CTR than in the Offs (30.5% vs 13%, $p = 0.0001$) or the AMI group (21.1%, $p = 0.016$; OR = 0.609). The A allele was significantly less frequent in the CTR than in the Offs (69% vs 87.7%, $p = 0.0001$) or the AMI group (78.9%, $p = 0.012$; OR = 1.674). There were also some differences between the Offs and the AMI group: the GG genotype was more frequent in the latter (21.1% vs 13%, $p = 0.037$; OR = 1.795); there was a higher percentage of A carriers among the Offs (87.7% vs 78.9%, $p = 0.02$), in whom the frequency of the AA genotype was also higher (35.1% vs AMI = 25.7%, $p = 0.043$).

Table 4 shows IFN- γ genotype and allele distribution between Offs, AMI and CTR groups.

Table 4 SNP IFN- γ (rs 2430561) genotype distribution and allele frequency from Offs, CTR and AMI.

IFN- γ	TT (n) %	TA (n) %	AA (n) %	T carr (n) %	A carr (n) %
Offs (n = 153)	(24) 15.7	(88) 57.5	(41) 26.8	(112) 73.2	(129) 84.3
CTR (n = 268)	(82) 30.6	(111) 41.4	(75) 28	(193) 72.0	(186) 69.4
AMI (n = 240)	(58) 24.2	(96) 40	(86) 35.8	(154) 64.2	(182) 75.8
Offs vs CTR	$\chi^2 = 13.990$, $p = 0.001$; TT carriers vs non-TT carriers: $\chi^2 = 11.070$, $p = 0.001$; A carriers $\chi^2 = 11.495$, $p = 0.001$				
AMI vs CTR	$\chi^2 = 4.423$, $p = 0.110$				
Offs vs AMI	$\chi^2 = 11.704$, $p = 0.003$; TT carriers vs non-TT carriers: $\chi^2 = 4.380$, $p = 0.036$; OR = 1.761 (CI: 1.032-3.002); A carriers $\chi^2 = 4.070$, $p = 0.044$; OR = 0.584 (CI: 0.345-0.988).				

The TT genotype was more frequent in the CTR than in the Offs (30.6% vs 15.7%, $p = 0.001$), whereas the percentage of A carriers was higher among the Offs than the CTR (84.3% vs 69.4%, $p = 0.001$). No difference in the distribution of the IFN- γ polymorphism between the AMI group and the CTR was detected, but the frequency of the TT genotype was slightly higher in the AMI group than in Offs (24.2% vs 15.7%, $p = 0.036$; OR = 1.761) and the A allele was less frequent in the AMI group than Offs (75.8% vs 84.3%, $p = 0.044$; OR = 0.584). The SNPs in the promoter region of the ACT, HMGCR and IL-1 β genes were also investigated, but not significant difference in allele and genotype frequencies between the groups was found (data not shown).

Association between the triple genotype and cardiovascular risk.

The concomitant presence of the CC genotype of VEGF, the A allele of IL-10 and the A allele of IFN- γ between Offs, AMI and CTR groups (that we called “triple genotype”) was also determined and resulted to be associated with an increased risk of AMI, as shown in Table 5.

Table 5 Concomitant presence of the triple genotype in Offs, CTR and AMI

Triple genotype	Carriers (n) %	Non-carriers (n) %
Offs (n = 153)	(71) 46.4	(82) 53.6
CTR (n = 301)	(52) 17.3	(248) 82.7
AMI (n = 239)	(76) 31.8	(163) 68.2
Offs vs CTR	$\chi^2 = 43.295$, $p = 0.0001$	
AMI vs CTR	$\chi^2 = 15.372$, $p = 0.0001$; OR = 2.224 (CI: 1.484-3.332)	

This “triple genotype” was more frequent in the Offs (46.4%) and the AMI (31.8%) than in CTR (17.3%), and the differences were highly statistically significant (Offs vs CTR: $p = 0.0001$, OR = 4.129; AMI vs CTR: $p = 0.0001$, OR = 2.224).

Body mass index (BMI) and blood lipid profile.

Data regarding BMI and serum lipid profile from Offs, CTR and IMA have been reported in Table 6.

Table 6 BMI values and blood lipid profiles from Offs, CTR and AMI.

		N°	Mean	St. Deviation	Post hoc statistics
BMI	Offs	154	27	4	Offs vs CTR*
	CTR	320	29	4	Offs vs IMA**
	IMA	265	27	4	CTR vs IMA*
Total cholesterol	Offs	148	216	43	Offs vs CTR*
	CTR	265	241	39	Offs vs IMA*
	IMA	209	199	39	CTR vs IMA*
HDL	Offs	148	60	17	Offs vs CTR**
	CTR	265	60	15	Offs vs IMA*
	IMA	209	47	16	CTR vs IMA*
LDL	Offs	137	132	36	Offs vs CTR*
	CTR	241	157	34	Offs vs IMA**
	IMA	189	127	38	CTR vs IMA*
Triglycerides	Offs	148	111	64	Offs vs CTR**
	CTR	265	122	70	Offs vs IMA*
	IMA	209	140	117	CTR vs IMA*
VLDL	Offs	148	22	13	Offs vs CTR**
	CTR	265	24	14	Offs vs IMA*
	IMA	209	28	23	CTR vs IMA*

*p = 0.0001.

**p ≥ 0.05; not statistically significant.

BMI values of Offs and IMA were slightly increased (27 ± 4) as compared with ideal age matched reference value. Moreover, BMI from our CTR group was higher than Offs and IMA, however this difference may be mainly ascribed to the older age of CTR. Lipid profile from the Offs population was substantially in the normal range for their age cohort. Once again CTR

population showed slightly increased blood levels of total cholesterol, LDL and triglycerides as expected according their age range.

Comparison for BMI, total cholesterol, HDL, LDL, triglycerides and VDL values between Offs carriers and non carriers for the triple genotype was performed. The results were display in Table 7.

Table 7 BMI and blood lipid parameters in Offs carriers or non carriers of the triple genotype.

	Triple genotype Offs	N°	Mean	St. Deviation	statistics
BMI	Carriers	71	26	4	not significant
	Not carriers	82	27	4	
Total cholesterol	Carriers	66	215	35	not significant
	Not carriers	81	215	47	
HDL	Carriers	66	60	14	not significant
	Not carriers	81	60	18	
LDL	Carriers	64	134	31	not significant
	Not carriers	72	130	38	
Triglycerides	Carriers	66	100	53	not significant
	Not carriers	81	120	74	
VLDL	Carriers	66	20	11	not significant
	Not carriers	81	24	15	

As shown in Table 7, no statistical difference for BMI, total cholesterol, HDL, LDL, triglycerides and VDL values between Offs carriers and non carriers for the triple genotype was detected.

Prevalence of CVE after 24 years of follow up.

The prevalence rates of a history of ischemic heart disease (AMI or angina pectoris), stroke, ischemic heart disease plus stroke, hypertension, diabetes and smoking in 154 Offs at the beginning (age 23–35 years; Table 8, panel A) and at the end of the follow up period (age 50– 60 years; Table 8, panel B) were compared with those of gender and age matched subjects from the MONICA Brianza population.

Table 8 Cardiovascular events in Controls and Offs during a 24 years follow up.

	panel A									
	Male					Female				
	MONICA-Brianza		Offs		P	MONICA-Brianza		Offs		P
	Age 25-35		(1984)		†	Age 25-35		(1984)		†
	N	%	N	%		N	%	N	%	
Subjects	727	-	84	-	-	768	-	70	-	-
Ischemic heart disease*	3	0.4	1	1.2	‡	2	0.3	0	0	‡
Stroke	1	0.1	0	0	‡	0	0	0	0	‡
Heart disease/stroke	4	0.6	1	1.2	‡	2	0.3	0	0	‡
Hypertension	72	9.9	4	4.8	‡	63	8.2	2	2.9	‡
Diabetes mellitus	2	0.3	0	0	‡	2	0.3	1	1.4	‡
Smoking	292	40.2	29	34.5	‡	240	31.3	27	38.6	‡

	panel B									
	Male					Female				
	MONICA-Brianza		Offs		P	MONICA-Brianza		Offs		P
	Age 50-60		(2008)		†	Age 50-60		(2008)		†
	N	%	N	%		N	%	N	%	
Subjects	970	-	84	-	-	967	-	70	-	-
Ischemic heart disease*	38	3.9	9	10.7	0.01	16	1.7	1	1.4	‡
Stroke	6	0.6	4	4.8	0.01	3	0.3	0	0	‡
Heart disease/stroke	44	4.5	13	15.5	0.0001	19	2	1	1.4	‡
Hypertension	268	27.6	29	34.5	‡	350	36.2	20	28.6	‡
Diabetes mellitus	39	4	8	9.5	0.05	30	3.1	5	7.1	‡
Smoking	354	36.5	17	20.2	0.003	145	15	13	18.6	‡

*Myocardial infarction or angina pectoris.

† Fisher's exact test.

‡ Not significant ($p > 0.05$).

No difference in the event rates was present between the two populations at the beginning of the follow up period. On the contrary, at the end of the follow up the prevalence of ischemic heart disease among the male Offs was three times higher, that of stroke was eight times higher, that of stroke and ischemic heart disease was three times higher, and that of diabetes was twice as high. However, there was no increased prevalence of CVE among female Offs during the same period.

Detection of EBV and HHV-6 DNA

Data regarding primers, PCR conditions and assay sensitivity for the detection of CMV, EBV, and HHV-6 nucleic acid positivity are reported in Table 9.

Table 9 Set of herpes virus primers used in quantitative real-time PCR (qPCR) reactions

Virus	Primer sets (nucleotide sequence 5' → 3')	Amplicon size/protein	PCR cycle conditions	LOD
EBV	GCCAGAGGTAAGTGGACTTTAAT GAGGGGACCCTGAGACGGGT	96 bp	50 °C: 2 min, 95 °C: 15 min, 45 cycles of 94 °C: 2 min, 60 °C: 30 s, 72 °C: 30 s, melt curve 65 °C to 95 °C, 4 °C.	33 copies/reaction
HHV-6	TCCATTATTTGGCCGCATTCGT TGTTAGGATATACCGATGTGCGT	173 bp	50 °C: 2 min, 95 °C: 15 min, 45 cycles of 94 °C: 2 min, 60 °C: 30 s, 72 °C: 30 s, melt curve 65 °C to 95 °C, 4 °C.	33 copies/reaction

DNA from CMP, Offs and CTR PBL samples were analyzed by qRT-PCR for the presence of HHV6 and EBV.

In Table 10, data regarding EBV positivity in PBL samples are shown.

Table 10. Presence or absence of Epstein Barr virus (EBV) DNA in CMP, CTR and Offs

EBV	positive	negative
CMP (n=396) age 70±11	n=250 63.1%	n=146 36.9%
CTR (n=136) age 72±5	n=79 58.1%	n=57 41.9%
Offs (n=109) age 56±7	n=52 47.7%	n=57 62.3%
CMP vs CTR	$\chi^2=1.091$ $p=0.296$ Log Reg for Age $p=0.952$	
CMP vs Offs	$\chi^2=8.460$ $p=0.004$ OR=1.877 CI(1.224-2.879) Log Reg for Age $p=0.432$	
Offs vs CTR	$\chi^2=2.621$ $p=0.105$ Log Reg for Age $p=0.483$	

In all samples, 63.1% of CMP patients, 58.1% of CTR and 47.7% of Offs were positive for EBV DNA. No differences between CMP and CTR and between Offs and CTR were found (CMP vs CTR $\chi^2=1.091$; $p=0.296$; Offs vs CTR $\chi^2=2.621$; $p=0.105$). Instead, statistically significant differences between CMP and Offs were found (CMP vs Offs $\chi^2=8.460$; $p=0.004$; OR=1.877; CI (1.224-2.879)).

A correction for age by a logistic regression analysis between CMP, CTR and Offs was applied but no statistically significant differences were found (CMP vs CTR $p=0.952$; CMP vs Offs $p=0.432$; Offs vs CTR $p=0.483$).

CMP patients were stratified in patients with AMI (CMP AMI) or Angina (aCMP). Data regarding EBV positivity in CMP AMI and aCMP, Offs and CTR are shown in Table 11.

Table 11. Presence or absence of Epstein Barr virus (EBV) DNA in CMP AMI, aCMP, CTR and Offs.

EBV	positive		negative	
CMP AMI (n=182)	n=109	59.9%	n=73	40.1%
aCMP (n=214)	n=141	65.9%	n=73	34.1%
CTR (n=136)	n=79	58.1%	n=57	41.9%
Offs (n=109)	n=52	47.7%	n=57	62.3%
CMP AMI vs aCMP	$\chi^2=1.520$ $p=0.218$			
CMP AMI vs CTR	$\chi^2=0.080$ $p=0.777$			
aCMP vs Offs	$\chi^2=9.927$ $p=0.002$ OR=2.117 CI=1.323-3.388			
CMP AMI vs Offs	$\chi^2=3.936$ $p=0.047$ OR=1.622 CI=1.004-2.618			
aCMP vs CTR	$\chi^2=2.167$ $p=0.141$			

Among CMP patients, 59.9% of CMP AMI and 65.9% of aCMP were positive for EBV DNA and no differences regarding the presence of EBV in CMP group were present. Statistically significant differences were observed between aCMP and Offs and CMP AMI and Offs (aCMP vs Offs: $\chi^2=9.927$; $p=0.002$; OR=2.117; CI=1.323-3.388. CMP AMI vs Offs: $\chi^2=3.936$; $p=0.047$; OR=1.622; CI=1.004-2.618).

EBV positive Offs and CTR subjects were stratified according to the presence of triple genotype carriers or not carriers. Data regarding EBV positivity in triple genotype carriers or not carriers Offs and CTR are shown in Table 12.

Table 12. Presence or absence of Epstein Barr Virus (EBV) in Triple Genotype Carriers and Non Carriers Offs and CTR.

EBV positive					
Triple genotype		Carriers		Non carriers	
Offs	(n=52)	n=22	42.3%	n=30	57.7%
CTR	(n=75)	n=12	16%	n=57	84%
Offs vs CTR $\chi^2=10.841$ $p=0.001$					
OR=3.850 CI (1.684-8.802)					

As shown in Table 12, 42.3% of Offs versus 16% of CTR were triple genotype carriers and EBV positive and the differences are statistically different (Offs vs CTR: $\chi^2=10.841$; $p=0.001$; OR=3.850; CI (1.684-8.802). The presence/absence of the triple genotype in Offs and CTR

subjects appeared to influence EBV positivity.

In Table 13, data regarding HHV-6 positivity in PBL samples are shown.

Table 13. Presence or absence of Herpes Virus-6 (HHV-6) DNA in CMP, CTR and Offs

HHV6	positive		negative	
CMP (n=380) Age 70±11	n=161	42.4%	n=219	57.6%
CTR (n=77) Age 72±5	n=49	63.6%	n=28	36.4%
Offs (n=135) Age 56±7	n=47	34.8%	n=88	65.2%
CMP vs CTR	$\chi^2=11.661$ p=0.001 OR=0.421 CI (0.253-0.697)			
	Log Reg for Age p=0.312			
CMP vs Offs	$\chi^2=2.361$ p=0.124			
	Log Reg for Age p=0.466			
Offs vs CTR	$\chi^2=16.439$ p=0.0001 OR=0.305 CI (0.170-0.547)			
	Log Reg for Age p=0.558			

In all samples, 42.4% of CMP patients, 63.6% of CTR and 56% of Offs were positive for HHV-6 DNA. At first glance, the HHV-6 positivity seemed to have a correlation with age. A logistic regression was applied to correct analysis for age between CMP, CTR and Offs but no statistically significant differences were found (CMP vs CTR: Log Reg for Age p=0.312; CMP vs Offs: Log Reg for Age p=0.466; Offs vs CTR: Log Reg for Age p=0.558).

CMP patients were stratified in AMI patients (CMP AMI) and angina (aCMP). Data regarding HHV-6 positivity in CMP AMI and aCMP, Offs and CTR are shown in Table 14.

Table 14. Presence or absence of Herpes Virus-6 (HHV-6) DNA in CMP AMI, aCMP, CTR and Offs

HHV6	positive		negative	
CMP AMI(n=177)	n=73	41.2%	n=104	58.8%
aCMP (n=203)	n=88	43.3%	n=115	56.7%
CTR (n=77)	n=49	63.6%	n=28	36.4%
Offs (n=135)	n=47	34.8%	n=88	65.2%
CMP AMI vs aCMP $\chi^2=0.172$ p=0.678				
CMP AMI vs CTR $\chi^2=10.534$ p=0.001 OR=0.405 CI= (0.233-0.704)				
aCMP vs Offs $\chi^2=2.462$ p=0.117				
CMP AMI vs Offs $\chi^2=1.431$ p=0.232				
aCMP vs CTR $\chi^2=9.194$ p=0.002 OR=0.437 CI= (0.255-0.751)				

Among investigated subjects, 41.2% of CMP AMI patients, 43.3% of aCMP, 63.6% of CTR and 34.8% of Offs were positive for HHV-6 DNA. No differences regarding the presence of HHV-6 between the two groups of CMP or between CMP AMI, aCMP, Offs and CTR are present. (CMP AMI vs aCMP: $\chi^2=0.172$; p=0.678; CMP AMI vs CTR: $\chi^2=10.534$; p=0.001; OR=0.405; CI=(0.233-0.704); aCMP vs Offs: $\chi^2=2.462$; p=0.117; CMP AMI vs Offs: $\chi^2=1.431$; p=0.232; aCMP vs CTR: $\chi^2=9.194$; p=0.002; OR=0.437; CI=0.255-0.751).

HHV-6 positive Offs and CTR were stratified according to the presence of triple genotype carriers or not carriers. Data regarding HHV-6 positivity in triple genotype carriers or not carriers Offs and CTR are shown in Table 15.

Table 15. Presence or absence of Herpes Virus-6 (HHV-6) in triple genotype carriers and non carriers Offs and CTR.

HHV6 positive					
Triple genotype		Carriers		Non carriers	
Offs	(n=47)	n=20	42.6%	n=27	57.4%
CTR	(n=48)	n=8	16.7%	n=40	83.3%
Offs vs CTR $\chi^2=7.656$ $p=0.006$					
OR=3.704 CI (1.426-9.617)					

In the analyzed groups, 42.6% of Offs and 16.7% of CTR were triple genotype carrier .The presence/absence of the triple genotype in Offs and CTR subjects appeared to affect HHV-6 positivity and the differences were statistically different (Offs vs CTR: $\chi^2=7.656$; $p=0.006$; OR=3.704 CI (1.426-9.617)).

DISCUSSION

The multiple pathogenetic pathways leading to AMI include classical and new risk factors and multiple genetic traits and environmental factors such as infections microorganisms might be associated with the disease. Recent genome-wide association (GWA) studies have contributed substantially to the discovery of new SNPs associated with CHD and AMI (Patel RS et al., 2011) but their clinical relevance is still unclear. In fact, a single gene variant can make a limited contribution to the total genetic load of AMI, and both common and rare gene polymorphisms may differentially affect susceptibility to the disease. These factors may also partially explain the contradictory results of genetic association studies using the candidate gene approach in AMI case/control studies (Kullo IJ et al., 2007; Hamsten A et al., 2008; Chiappelli M et al., 2005).

Familiarity and AMI

Parental history is a widely accepted risk factor for offspring of parents with CVE and AMI, although the mechanisms remain unclear. For instance, Offs with positive parental history for CVE (<65 years) resulted at higher risk of incident CVE compared with Offs with negative parental history. Despite Offs with positive parental history showed classical risk factors such as increased blood pressure, total and high-density lipoprotein cholesterol, CRP levels and decreased physical activity, less than 15% of the excess risk was explained through these risk factors. (Hamer M et al., 2009).

Family history of AMI, stroke, and related risk factors in first-degree relatives was studied. By looking at the extent to which parental history was associated with affected siblings within disease category. 15.7% cases of acute coronary syndromes occurred in families with ≥ 2 affected first-degree relatives compared with 5.1% strokes. Therefore, heritability of coronary events appeared to be greater than that of cerebral events and AMI was more likely to cluster in families than stroke (Banerjee A et al., 2011).

It was recently shown the presence of a relation between a family history of heart attack and the occurrence of early AMI in young women. In fact, the rate of AMI among women first-degree

relatives of AMI cases was twice as high as among first-degree relatives of controls. These findings suggested that family history of AMI was positively associated with the risk of early AMI in women (Friedlander Y et al., 2001).

It is important to know that subjects with an affected parent have a two-fold greater risk of CHD than those without a positive family history (Rumboldt M et al., 2003; Lloyd-Jones DM et al., 2004; Murabito JM et al., 2005). Genetic studies from children of parents with CVD have shown that genetic variations in the promoter region of the APOA1 gene were associated with differences in serum ApoA1 and HDL levels in unaffected subjects; this association was also influenced by gender and a family history of AMI (Talmud PJ et al., 1994).

A family history and a still largely undefined genetic background greatly influence the early clinical manifestation of AMI and CVD. Therefore, we investigated SNPs in genes with a regulatory effect on inflammatory responses and cholesterol metabolism as possible genetic markers of an increased risk of CVD in Offs with a positive history of AMI.

SNPs in the VEGF, IL-10 and IFN- γ genes were differently distributed in Offs and CTR. We chose elderly CTR from a longitudinal population study because they did not have a history of CVD, and did not experience an AMI or have any other CVD before and during the five years follow-up. It is important to note that 45% of the Offs in our investigation had a father who suffered an AMI before the age of 56 years. The genetic make-up of the Offs overlapped that of the unrelated population of patients with a clinical diagnosis of sporadic AMI (control disease).

Genetic variations and AMI

It has been suggested that the CC genotype of the VEGF gene, together with two other SNPs in the promoter region of this gene, increases VEGF gene expression in human myoblasts (Prior SJ et al., 2006) and the production of the cognate protein in human peripheral blood lymphocytes activated by lipopolisaccaride (Mohammadi M e al., 2009). Therefore, subjects with the VEGF CC genotype may produce increased levels of VEGF protein which, by deregulating angiogenesis, may lead to an increased risk of CVE.

These data were supported from previously published data regarding the functional relevance of IL-10 SNPs. An initial study found that the IL-10 A allele is associated with a two-fold increase in transcriptional activity in B cell lines (Rees LE et al., 2002), but it has been subsequently reported that the A allele is associated with a reduction in the IL-10 secretion of activated peripheral blood lymphocytes (Van der Linde K et al., 2003; Stanilova SA et al., 2006; Aborsangaya KB et al., 2007).

This finding suggest that the suppression of inflammation may be impaired in carriers of the IL-10 -1082 A allele and that an impaired regulation of inflammatory responses in Offs with one or two copies of the IL-10 A allele may increase the risk of CVE.

A decreased release of IFN- γ may negatively affect the coordination of immune responses in vessel walls, accelerate atherogenesis, and increase the risk of CVE in Offs with the IFN- γ A allele. Our findings showing an increased frequency of the +874 A allele in Offs are in accordance with another study reporting that patients with the IFN- γ +874 AA genotype and idiopathic dilated cardiomyopathy showed a worse prognosis and an adverse outcome (Adamopoulos S et al., 2011). Furthermore, patients with dilated cardiomyopathy showed an impaired activation of CD4T cells by IFN- γ ascribed to a decreased production of this cytokine (Lindberg E et al., 2010).

The concomitant presence of the CC genotype of the VEGF gene, the A allele of the IL-10 gene, and the A allele of the IFN- γ gene was more frequent in our AMI group, and significantly increased the risk of the disease. It is interesting to note that this triple genotype profile was found in 46% of Offs and led to a presumptive high risk of CVE (OR = 4.129). This observation supports the notion that Offs with a positive parental history are at high risk of CVE and the pro-inflammatory genetic signature may be part of the complex genetic background influencing AMI risk. Offs were followed up for a 24 years period and the occurrence of CVE registered.

A control group of comparable age from the WHO-MONICA-Brianza study to compare the CVE prevalence during the 24 years follow-up of our Offs was also used.

The potential increased risk of CVE predicted by pro-inflammatory genetic signature was

partially confirmed by data from the 24-year follow-up, which clearly revealed a significant increase in CVE in Offs. Furthermore, gender and age were conditions with a strong influence on CVE, since increased prevalence of CVE was observed only in male Offs (Table 8). The possible relationship between the presence of the triple genotype and CVE manifestation in Offs could not be assessed, since the limited number of Offs positive for CVE. Further follow up of these subjects may clarify this topic.

BMI from Offs was slightly increased when compared with the ideal age value and CTR selected for this investigation showed an increased BMI as expected since their advanced age. Cholesterol blood profile data from Offs group were in the normal range for their age cohort. Among Offs only 33 subjects used for limited time periods statins. Therefore, statins use showed very limited influence upon the lipid profile data from Offs population. Furthermore, since the presence of the triple genotype/allele signature did not affect BMI or the blood levels of total cholesterol, HDL, LDL or triglycerides, we suggest that it might affect the incidence of CVE by mechanisms that are partially independent of those affected by the classic risk factors of CVD.

Virus infections and AMI

EBV infects more than 95% of human beings within the first years of life. The virus causes acute infections (mononucleosis) in a minority of immune-competent subjects, whereas the majority develops a lifelong asymptomatic infection, and the virus remains latent in memory B-lymphocytes. EBV is also involved in the development of several diseases such as Burkitt lymphoma, Hodgkin lymphoma, and nasopharyngeal carcinoma (Kutok JL et al., 2006). Moreover, EBV seems to be involved in the pathogenesis of various neurological diseases, such as encephalitis, neuritis, myelitis, cerebellitis, acute disseminated encephalomyelitis, central nervous system (CNS) lymphoma in patients with human immunodeficiency virus (HIV) infection (Kleines M et al., 2011) and multiple sclerosis (Lassmann H et al., 2011).

HHV-6 is a neurotropic virus and has been associated with multiple neurological diseases and disease conditions including seizures, encephalitis, mesial temporal lobe epilepsy, and multiple

sclerosis (Yao K et al., 2010). HHV-6 has been found in a higher proportion in AD brain than in age-matched controls (Lin WR et al., 2002).

Evidence for EV, adenovirus, and CMV, parvovirus B19, EBV, HHV-6 association with dilated cardiomyopathy has been reported (Pankuweit S et al., 2010; Al-Ghamdi A, 2012). The levels of CMV- and EBV- specific antibodies were found elevated among patients with CVD and the presence of CMV-specific IgG was associated with development of the disease. The seroprevalence of anti-H. pylori, anti-CMV and anti-HSV-1 antibodies in AMI and unstable angina groups was significantly higher than those of the to control group (Jafarzadeh A 2011). Genomic sequences of EBV and HHV-8 DNA in atherosclerotic lesion was found, thus suggesting that the viruses could play a role in atherogenesis (Tremolada S et al., 2011; Kaklikkaya I et al., 2010). Adv3 and HHV6 sequences coexisted in one case with inflammatory cardiomyopathy (Tátrai E et al., 2010).

The seropositivity to CMV, EBV, or HHV-6 in general population is very high. Moreover, we were also interested in the immune response of the host to infections, since with aging the immune system undergoes significant changes. This process has been called immunosenescence and may lead to an increased susceptibility to develop not only infectious diseases but also CVD, AD, osteoporosis, autoimmunity, and cancer in elderly (Lang PO et al., 2012).

However, the data regarding seroprevalence and genomic positivity for EBV and HHV-6 viruses in CVD and AMI remain controversial. In fact, it is difficult to find a linear association of serological viral positivity with AMI, since of the high seroprevalence in the elderly population.

Our investigations regarding EBV and HHV-6 DNA presence in PBL samples suggest an association of EBV and HHV-6 DNA with CVD and AMI.

In fact, we found that a high proportion of CMP patients had EBV positive blood leukocytes, since 63% of CMP, 58% CTR and 47.7% of Offs were EBV positive. Statistically significant differences between CMP and Offs were detected. When CMP were stratified in AMI CMP and aCMP statistically significant differences remained among AMI CMP, aCMP and Offs. Among the analyzed subjects 42.3% of Offs versus 16% of CTR were triple genotype carriers. The

presence of the triple genotype was increased in Offs and this group showed also a high positivity for EBV. These findings suggest that the presence of EBV DNA in the PBL might contribute to the risk for CVD. EBV positivity was also high in elderly CTR. The immunosenescence process of elderly may explain the increased EBV positivity.

A higher proportion of elderly CTR than CMP and Offs was positive for HHV-6 (63.6% vs 42.4 and 43.8%, respectively). These differences remained when CTR were compared with the two subgroups AMI CMP and aCMP. These findings suggest that senescence of immune responses might influence both virus subclinical infections and CVD. Whether EBV and HHV-6 persistence may independently affect CVD clinical history remains an open question. Between the analyzed subjects 42.6% of Offs versus 16.7% of CTR were triple genotype carriers. Offs with the triple genotype showed also an increased HHV-6 positivity.

These findings showing an association between elevated EBV and HHV-6 DNA positivity, suggesting a role of chronic virus infections in CVD by interfering with the efficiency of immune responses.

CONCLUSIONS

The concomitant presence of the CC genotype of VEGF, the A allele of IL-10 and the A allele of IFN- γ resulted to be associated with an increased risk of AMI and was more frequent among Offs with a positive parental history of AMI. Therefore, genetic variations impairing the immune functions appear to be associated with the pathogenesis of AMI. Inflammatory genes, gender and age influence an accelerated aging of cardiovascular system and selected genes with immune regulatory functions are part of the complex genetic background contributing to familiarity for cardiovascular diseases.

The triple genotype seems to be associated also with the presence of EBV and HHV-6 DNA in Offs.

These preliminary results need to be interpreted with caution until they can be replicated in a larger case-control study. However, our findings support the notion that persistent cycles of latency and reactivation phases of these viruses by stressing the systemic immune responses might induce altered inflammatory processes that, in turn, may accelerate cardiovascular aging and increase the prevalence of CVD. Therefore, EBV and HHV-6 might be considered environmental risk factors for CVD and AMI. Vaccination or antiviral therapy might in the mean future decrease the incidence and prevalence of CVD in the elderly.

Further studies are needed to clarify the primary or secondary role of herpes virus infection in CVD and AMI. In particular, improved methodology focused on investigating viral latency and its effect on immune system might possibly help to better elucidate the role of these pathogens in CVD pathogenesis.

REFERENZE

1. Aborsangaya KB, Dembinski I, Khatkar S, Alphonse MP, Nickerson P, Rempel JD. Impact of aboriginal ethnicity on HCV core-induced IL-10 synthesis: interaction with IL-10 gene polymorphisms. *Hepatology* 2007, 45:623–30.
2. Adamopoulos S, Kolokathis F, Gkouziouta A, Georgiadou P, Chaidaroglou A, Karavolias GK, Degiannis D, Voudris V, Kremastinos DT. Cytokine gene polymorphisms are associated with markers of disease severity and prognosis in patients with idiopathic dilated cardiomyopathy. *Cytokine* 2011, 54:68–73.
3. Adams ER, Nolan VG, Andersen SL, Perls TT, Terry DF. Centenarian offspring: start healthier and stay healthier. *J Am Geriatr Soc* 2008, 56:2089–92.
4. Al-Ghamdi A. Role of herpes simplex virus-1, cytomegalovirus and Epstein-Barr virus in atherosclerosis. *Pak J Pharm Sci* 2012, 25(1):89-97.
5. Allen N, Berry JD, Ning H, Van Horn L, Dyer A, Lloyd-Jones DM. Impact of blood pressure and blood pressure change during middle age on the remaining lifetime risk for cardiovascular disease: the cardiovascular lifetime risk pooling project. *Circulation* 2012, 125(1):37-44.
6. Andreotti F, Porto I, Crea F, Maseri A. Inflammatory gene polymorphisms and ischaemic heart disease: review of population association studies. *Heart* 2002, 87:107–12.
7. Andrié RP, Bauriedel G, Tuleta I, Braun P, Nickenig G, Skowasch D. Impact of intimal pathogen burden in acute coronary syndromes--correlation with inflammation, thrombosis, and autoimmunity. *Cardiovasc Pathol* 2010, 19(6):e205-10.
8. Andrié R, Braun P, Heinrich KW, Lüderitz B, Bauriedel G. Prevalence of intimal pathogen burden in acute coronary syndromes. *Z Kardiol.* 2003, 92(8):641-9.
9. Arroyo-Espliguero R, Avanzas P, Cosín-Sales J, Aldama G, Pizzi C, Kaski JC. C-reactive protein elevation and disease activity in patients with coronary artery disease. *Eur Heart J* 2004, 25(5):401-8.
10. Banerjee A, Silver LE, Heneghan C, Welch SJ, Mehta Z, Banning AP, Rothwell PM. Relative familial clustering of cerebral versus coronary ischemic events. *Circ Cardiovasc Genet*

2011, 4(4):390-6.

11. Barra S, Gaeta G, Cuomo S, Guarini P, Foglia MC, Capozzi G, Materazzi C, Trevisan M. Early increase of carotid intima-media thickness in children with parental history of premature myocardial infarction. *Heart* 2009, 95:642–5.

12. Bayram A, Erdoğan MB, Ekşi F, Yamak B. Demonstration of *Chlamydia pneumoniae*, *Mycoplasma pneumoniae*, Cytomegalovirus, and Epstein-Barr virus in atherosclerotic coronary arteries, nonrheumatic calcific aortic and rheumatic stenotic mitral valves by polymerase chain reaction. *Anadolu Kardiyol Derg* 2011,11(3):237-43.

13. Bennet AM, Di Angelantonio E, Ye Z, Wensley F, Dahlin A, Ahlbom A, Keavney B, Collins R, Wiman B, de Faire U, Danesh J. Association of apolipoprotein E genotypes with lipid levels and coronary risk. *JAMA* 2007, 298(11):1300-11.

14. Braunwald E. Shattuck Lecture-cardiovascular medicine at the turn of the millenium: triumphs, concerns and opportunities. *N Engl J Med* 1997, 337:1360–9.

15. Candore G, Balistreri CR, Grimaldi MP, Listì F, Vasto S, Caruso M, Caimi G, Hoffmann E, Colonna-Romano G, Lio D, Paolisso G, Franceschi C, Caruso C. Opposite role of pro-inflammatory alleles in acute myocardial infarction and longevity: results of studies performed in a Sicilian population. *Ann N Y Acad Sci* 2006,1067:270-5.

16. Chiappelli M, Tampieri C, Tumini E, Porcellini E, Caldarera CM, Nanni S, Branzi A, Lio D, Caruso M, Hoffmann E, Caruso C, Licastro F. Interleukin-6 gene polymorphism is an age-dependent risk factor for myocardial infarction in men. *Int J Immunogenet* 2005, 32:349–53.

17. Chobanian AV , Bakris GL , Black HR , Cushman WC , Green LA , Izzo JL Jr , Jones DW, Materson BJ, Oparil S, Wright JT Jr , Roccella EJ; National Heart, Lung, and Blood Institute Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure; National High Blood Pressure Education Program Coordinating Committee. The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. *JAMA* 2003, 289(19):2560-72.

18. Das SR, Alexander KP, Chen AY, Powell-Wiley TM, Diercks DB, Peterson ED, Roe MT, de

Lemos JA. . Impact of Body Weight and Extreme Obesity on the Presentation, Treatment, and In-Hospital Outcomes of 50,149 Patients With ST-Segment Elevation Myocardial Infarction Results From the NCDR (National Cardiovascular Data Registry). *J Am Coll Cardiol* 2011, 58(25):2642-50.

19. De Gennaro L, Brunetti ND, Montrone D, De Rosa F, Cuculo A, Di Biase M. Subacute inflammatory activation in subjects with acute coronary syndrome and left ventricular dysfunction.

Inflammation 2012, 35(1):363-70.

20. De Lemos JA, Morrow DA, Sabatine MS, Murphy SA, Gibson CM, Antman EM, McCabe CH, Cannon CP, Braunwald E. Association between plasma levels of monocyte chemoattractant protein-1 and long-term clinical outcomes in patients with acute coronary syndromes. *Circulation* 2003, 107(5):690-5.

21. Dong JY, Zhang YH, Qin LQ. Erectile dysfunction and risk of cardiovascular disease meta-analysis of prospective cohort studies. *J Am Coll Cardiol* 2011, 58(13):1378-85.

22. European Cardiovascular Disease Statistics 2012 Edition, p116. Retrieved 14 August 2013 <http://www.escardio.org/about/Documents/EUcardiovascular-disease-statistics-2012.pdf>.

23. Fahim MR, Halim SM, Kamel I. Tumor necrosis factor alpha in patients with acute myocardial infarction. *Egypt J Immunol* 2004, 11(1):31-7.

24. Ferrario M, Sega R, Chatenoud L, Mancia G, Mocarelli P, Crespi C, Cesana G, MONICA-Brianza Research Group. MONItoring of CArdiovascular diseases: Time trends of major coronary risk factors in a northern Italian population (1986–1994). How remarkable are socioeconomic differences in an industrialized low CHD incidence country? *Int J Epidemiol* 2001, 30:285–97.

25. Friedlander Y, Arbogast P, Schwartz SM, Marcovina SM, Austin MA, Rosendaal FR, Reiner AP, Psaty BM, Siscovick DS. Family history as a risk factor for early onset myocardial infarction in young women.. *Atherosclerosis* 2001, 156(1):201-7.

26. Galioto A, Dominguez LJ, Pineo A, Ferlisi A, Putignano E, Belvedere M, Costanza G,

- Barbagallo M. Cardiovascular risk factors in centenarians. *Exp Gerontol* 2008, 43:106–13.
27. Greenland P, Alpert JS, Beller GA, Benjamin EJ, Budoff MJ, Fayad ZA, Foster E, Hlatky MA, Hodgson JM, Kushner FG, Lauer MS, Shaw LJ, Smith SC Jr, Taylor AJ, Weintraub WS, Wenger NK, Jacobs AK; American College of Cardiology Foundation / American Heart Association Task Force on Practice Guidelines. ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults: executive summary: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. *J Am Coll Cardiol* 2010, 56(25):2182-99.
28. Grimaldi LM, Casadei VM, Ferri C, Veglia F, Licastro F, Annoni G, Biunno I, De Bellis G, Sorbi S, Mariani C, Canal N, Griffin WS, Franceschi M. Association of early-onset Alzheimer's disease with an interleukin-1alpha gene polymorphism. *Ann Neurol* 2000, 47:361–5.
29. Hamer M, Chida Y, Stamatakis E. The role of conventional and novel mechanisms in explaining increased risk of cardiovascular events in offspring with positive parental history. *J Hypertens* 2009, 27(10):1966-71.
30. Hamsten A, Eriksson P. Identifying the susceptibility genes for coronary artery disease: from hyperbole through doubt to cautious optimism. *J Intern Med* 2008, 263:538–52.
31. Hansson GK. Immune mechanisms in atherosclerosis. *Arteriosclerosis, Thrombosis and Vascular Biology* 2001, 21, 1876.
32. Havrylenko TI, Parkhomenko OM, Ryzhkova NO, Kozhukhov SM, Iakushko LV. Cytokine profile of mononuclear leukocytes in patients with myocardial infarction complicated by cardiac insufficiency. *Fiziol Zh* 2012, 58(6):23-8.
33. He M, van Dam RM, Rimm E, Hu FB, Qi L. Whole-grain, cereal fiber, bran, and germ intake and the risks of all-cause and cardiovascular disease-specific mortality among women with type 2 diabetes mellitus. *Circulation* 2010, 121(20):2162-8.
34. Hjemdahl P. Stress and the metabolic syndrome: an interesting but enigmatic association. *Circulation* 2002, 106(21):2634-6.
35. Howard BV, Rodriguez BL, Bennett PH, Harris MI, Hamman R, Kuller LH, Pearson TA,

Wylie-Rosett J. Prevention Conference VI: Diabetes and Cardiovascular disease: Writing Group I: epidemiology. *Circulation* 2002, 105(18):132-7.

36. Humphries SE, Luong LA, Ogg MS, Hawe E, Miller GJ. The interleukin-6 -174 G/C promoter polymorphism is associated with risk of coronary heart disease and systolic blood pressure in healthy men. *Eur Heart J* 2001, 22, 2243–52.

37. Huxley RR, Woodward M. Cigarette smoking as a risk factor for coronary heart disease in women compared with men: a systematic review and meta-analysis of prospective cohort studies. *Lancet* 2011, 378(9799):1297-305.

38. Incalcaterra E, Caruso M, Balistreri CR, Candore G, Lo Presti R, Hoffmann E, Caimi G. Role of genetic polymorphisms in myocardial infarction at young age. *Clin Hemorheol Microcirc* 2010, 46(4):291-8.

39. Ingelsson E, Lundholm C, Johansson AL, Altman D. Hysterectomy and risk of cardiovascular disease: a population-based cohort study. *Eur Heart J* 2011, 32(6):745-50.

40. Jafarzadeh A, Nemati M, Tahmasbi M, Ahmadi P, Rezayati MT, Sayadi AR The association between infection burden in Iranian patients with acute myocardial infarction and unstable angina. *Acta Med Indones* 2011, 43(2):105-11.

41. Jenny NS, Racy RP, Ogg MS, Luong IA, Kuller LH, Arnold AM, Sharrett AR, Humphries SE. In the elderly, interleukin-6 plasma levels and the -174 G>C polymorphism are associated with the development of cardiovascular disease. *Arterioscler Thromb Vasc Biol* 2002, 22:2066–71.

42. Kaklikkaya I, Kaklikkaya N, Birincioglu I, Buruk K, Turan N. Detection of human herpesvirus 6 DNA but not human herpesvirus 7 or 8 DNA in atherosclerotic and nonatherosclerotic vascular tissues. *Heart Surg Forum* 2010, 13(5):345-9.

43. Karpiński Ł, Plaksej R, Derzhko R, Orda A, Witkowska M . Serum levels of interleukin-6, interleukin-10 and C-reactive protein in patients with myocardial infarction treated with primary angioplasty during a 6-month follow-up. *Pol Arch Med Wewn* 2009, 119(3):115-21.

44. Kestenbaum B, Katz R, de Boer I, Hoofnagle A, Sarnak MJ, Shlipak MG, Jenny NS,

- Siscovick DS. Vitamin d, parathyroid hormone, and cardiovascular events among older adults. *J Am Coll Cardiol* 2011, 58(14):1433-41.
45. Kleines M, Schiefer J, Stienen A, Blaum M, Ritter K, Häusler M. Expanding the spectrum of neurological disease associated with Epstein-Barr virus activity. *Eur J Clin Microbiol Infect Dis* 2011, 30:1561-9.
46. Koch W, Wolferstetter H, Schatke A, Schömig A, Kastrati A. Interleukin 18 gene variation and risk of acute myocardial infarction. *Cytokine* 2011a, 56(3):786-91.
47. Koch W, Kastrati A, Böttiger C, Mehilli J, von Beckerath N, Schömig A. Interleukin-10 and tumor necrosis factor gene polymorphisms and risk of coronary artery disease and myocardial infarction. *Atherosclerosis*. 2011b, 159(1):137-44.
48. Konenkov VI, Shevchenko AV, Prokofev VF, Maksimov VN. Complex of genotypes of cytokines as a genetic factor of risk of development of myocardial infarction of in European population of Russia men. *Kardiologia* 2012, 52(7):22-9.
49. Kullo IJ, Ding K. Mechanisms of disease: The genetic basis of coronary heart disease. *Nat Clin Pract Cardiovasc Med* 2007, 4:558–69.
50. Kutok JL, Wang F. Spectrum of Epstein-Barr virus-associated diseases. *Annu Rev Pathol*, 2006, 1:375-404.
51. Lang PO, Govind S, Aspinall R. Reversing T cell immunosenescence: why, who, and how. *Age* 2012, 35:609-20.
52. LaRosa JC, Grundy SM, Waters DD, Shear C, Barter P, Fruchart JC, Gotto AM, Greten H, Kastelein JJ, Shepherd J, Wenger NK; Treating to New Targets (TNT) Investigators. . Intensive lipid lowering with atorvastatin in patients with stable coronary disease. *N Engl J Med* 2005, 352(14):1425-35.
53. Lassmann H, Niedobitek G, Aloisi F, Middeldorp JM, NeuroproMiSe EBV Working Group. Epstein-Barr virus in the multiple sclerosis brain: a controversial issuedreport on a focused workshop held in the Centre for Brain Research of the Medical University of Vienna, Austria. *Brain* 2011, 134: 2772-86.

54. Levenson J, Giral P, Razavian M, Garipey J, Simon A. Fibrinogen and silent atherosclerosis in subjects with cardiovascular risk factors. *Arterioscler Thromb Vasc Biol* 1995, 15:1263–83.
55. Levy D, Garrison RJ, Savage DD, Kannel WB, Castelli WP. Prognostic implications of echocardiographically determined left ventricular mass in the Framingham Heart Study. *N Engl J Med* 1990, 322(22):1561-6.
56. Licastro F, Chiappelli M, Caldarera CM, Tampieria C, Nannic S, Gallinac M, Branzi A. The concomitant presence of polymorphic alleles of interleukin-1b, interleukin-6 and apolipoprotein E is associated with an increased risk of myocardial infarction in elderly men Results from a pilot study, *Mech Ageing Dev* 2004, 125(8):575-9.
57. Licastro F, Chiapelli M, Caldarera CM, Caruso C, Lio D, Corder EH. Acute myocardial infarction and proinflammatory gene variants. *Ann N Y Acad Sci* 2007, 1119:227-42.
58. Licastro F, Chiappelli M, Porcellini E, Campo G, Buscema M, Grossi E, Garoia F, Ferrari R. Gene-gene and gene - clinical factors interaction in acute myocardial infarction: a new detailed risk chart. *Curr Pharm Des* 2010, 16(7):783-8.
59. Licastro F, Chiappelli M, Caldarera CM, Porcellini E, Carbone I, Caruso C, Lio D, Corder EH. Sharing pathogenetic mechanisms between acute myocardial infarction and Alzheimer's disease as shown by partially overlapping of gene variant profiles. *J Alzheimers Dis* 2011, 23(3):421-31.
60. Lin WR, Wozniak MA, Cooper RJ, Wilcock GK, Itzhaki RF. Herpesviruses in brain and Alzheimer's disease. *J. Pathol* 2002, 197:395-402.
61. Lindberg E, Andersson B, Hörnquist EH, Magnusson Y. Impaired activation of IFN-gamma + CD4+ T cells in peripheral blood of patients with dilated cardiomyopathy. *Cell Immunol* 2010, 263:224–9.
62. Lloyd-Jones DM, Nam BH, D'Agostino RB Sr, Levy D, Murabito JM, Wang TJ, Wilson PW, O'Donnell CJ. Parental cardiovascular disease as a risk factor for cardiovascular disease in middle-aged adults: a prospective study of parents and offspring. *JAMA* 2004, 291(18):2204-11.
63. Malkin CJ, Pugh PJ, Morris PD, Asif S, Jones TH, Channer KS. Low serum testosterone and

- increased mortality in men with coronary heart disease. *Heart* Nov 2010, 96(22):1821-5.
64. Magnoni M, Malnati M, Cristell N, Coli S, Russo D, Ruotolo G, Cianflone D, Alfieri O, Lusso P, Maseri A. Molecular study of human herpesvirus 6 and 8 involvement in coronary atherosclerosis and coronary instability. *J Med Virol* 2012, 84(12):1961-6.
65. Målarstig A, Eriksson P, Hamsten A, Lindahl B, Wallentin L, Siegbahn A. Raised interleukin-10 is an indicator of poor outcome and enhanced systemic inflammation in patients with acute coronary syndrome. *Heart*. 2008, 94(6):724-9.
66. Malorny B, Hoorfar J, Bunge C, Helmuth R. Multicenter validation of the analytical accuracy of Salmonella PCR: towards an international standard. *Appl Environ Microbiol* 2003, 69:290-6.
67. Micha R, Wallace SK, Mozaffarian D. Red and processed meat consumption and risk of incident coronary heart disease, stroke, and diabetes mellitus: a systematic review and meta-analysis. *Circulation* 2010, 121(21):2271-83.
68. Miller A, Kelly GS. Homocysteine metabolism: nutritional modulation and impact on health and disease. *Alter Med Rev* 1997, 2: 234–54.
69. Moe KT, Wong P. Current trends in diagnostic biomarkers of acute coronary syndrome. *Ann Acad Med Singap* 2010, 39(3): 210–5.
70. Mohammadi M, Bazrafshani MR, Day PJ, Ollier WE. Vascular endothelial growth factor production is regulated by gene polymorphisms. *Iran J Immunol* 2009, 6:119–29.
71. Murabito JM, Pencina MJ, Nam BH, D'Agostino RB Sr, Wang TJ, Lloyd-Jones D, Wilson PW, O'Donnell CJ. Sibling cardiovascular disease as a risk factor for cardiovascular disease in middle-aged adults. *JAMA* 2005, 294:3117–23.
72. Nazarenko GI, Skvortsova VI, Kleimenova EB, Konstantinova MV. The role of genetic predisposition in the development of cardiovascular diseases (myocardial infarction, ischemic stroke, unstable stenocardia) and its interaction with conventional risk factors *Zh Nevrol Psikhiatr Im S S Korsakova*. 2009, 109:19-25.
73. Nordmann AJ, Suter-Zimmermann K, Bucher HC, Shai I, Tuttle KR, Estruch R, Briel M

Meta-analysis comparing mediterranean to low-fat diets for modification of cardiovascular risk factors. *Am J Med* 2011, 124(9):841-81.

74. Pankuweit S, Maisch B. The heart in viral infections. *Internist* 2010, 51(7):836-43.

75. Patel RS, Ye S. Genetic determinants of coronary heart disease: new discoveries and insights from genome-wide association studies. *Heart* 2011, 97:1463–73.

76. Pesonen E, Andsberg E, Ohlin H, Puolakkainen M, Rautelin H, Sarna S, Persson K. Dual role of infections as risk factors for coronary heart disease. *Atherosclerosis* 2007, 192(2):370-5.

77. Prior SJ, Hagberg JM, Paton CM, Douglass LW, Brown MD, McLenithan JC, Roth SM. DNA sequence variation in the promoter region of the VEGF gene impacts VEGF gene expression and maximal oxygen consumption. *Am J Physiol Heart Circ Physiol* 2006, 290:1848–55.

78. Ravaglia G, Forti P, Maioli F, Orlanducci P, Sacchetti L, Flisi E, Dalmonte E, Martignani A, Cucinotta D, Cavalli G. Conselice study: a population based survey of brain aging in a municipality of the Emilia Romagna region: (A.U.S.L. Ravenna). Design and methods. *Arch Gerontol Geriatr Suppl* 2001, 7:313–24.

79. Ray KK, Camp NJ, Bennett CE, Francis SE, Crossman DC. Genetic variation at the interleukin-1 locus is a determinant of changes in soluble endothelial factors in patients with acute coronary syndromes. *Clin Sci* 2002, 103(3):303-10.

80. Rea TD, Heckbert SR, Kaplan RC, Smith NL, Lemaitre RN, Psaty BM. Smoking status and risk for recurrent coronary events after myocardial infarction. *Ann Intern Med* 2002, 137(6):494-500.

81. Rees LE, Wood NA, Gillespie KM, Lai KN, Gaston K, Mathieson PW. The interleukin-10-1082 G/A polymorphism: allele frequency in different populations and functional significance. *Cell Mol Life Sci* 2002, 59:560–9.

82. Rexrode KM, Carey VJ, Hennekens CH, Walters EE, Colditz GA, Stampfer MJ, Willett WC, Manson JE. Abdominal adiposity and coronary heart disease in women. *JAMA* 1998, 280(21):1843-8.

83. Ridker PM. Evaluating novel cardiovascular risk factors: can we better predict heart attacks? *Ann Int Med* 1999, 130:933–7;
84. Ridker PM, Rifai N, Pfeffer M, Sacks F, Lepage S, Braunwald E. Elevation of tumor necrosis factor- α and increased risk of recurrent coronary events after myocardial infarction. *Circulation* 2000a, 101: 2149-53.
85. Ridker PM, Rifai N, Stampfer MJ, Hennekens CH. Plasma concentration of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. *Circulation* 2000b, 101:1767–72.
86. Ridker PM, Danielson E, Fonseca FA, Genest J, Gotto AM Jr, Kastelein JJ, Koenig W, Libby P, Lorenzatti AJ, MacFadyen JG, Nordestgaard BG, Shepherd J, Willerson JT, Glynn RJ; JUPITER Study Group. Rosuvastatin to prevent vascular events in men and women with elevated C-reactive protein. *N Engl J Med* 2008, 359(21):2195-207
87. Rifai N, Ridker PM. Proposed cardiovascular risk assessment algorithm using high-sensitivity C-reactive protein and lipid screening. *Clin Chem* 2001, 47:28–30.
88. Rugina M, Caras I, Jurcut R, Jurcut C, Serbanescu F, Salageanu A, Apetrei E. Systemic inflammatory markers in patients with aortic sclerosis. *Roum Arch Microbiol Immunol* 2007, 66(1-2):10-6.
89. Rumboldt M, Rumboldt Z, Pesenti S. Premature parental heart attack is heralding elevated risk in their offspring. *Coll Antropol* 2003, 27(1):221-8.
90. Seshadri S, Beiser A, Pikula A, Himali JJ, Kelly-Hayes M, Debette S, De Stefano AL, Romero JR, Kase CS, Wolf PA. Parental occurrence of stroke and risk of stroke in their children: the Framingham study. *Circulation* 2010, 121:1304–12.
91. Stanilova SA, Miteva LD, Karakolev ZT, Stefanov CS. Interleukin-10-1082 promoter polymorphism in association with cytokine production and sepsis susceptibility. *Intensive Care Med* 2006, 32:260–6.
92. Talmud PJ, Ye S, Humphries SE. Polymorphism in the promoter region of the apolipoprotein AI gene associated with differences in apolipoprotein AI levels: the European

Atherosclerosis Research Study. *Genet Epidemiol* 1994, 11:265–80.

93. Tan J, Hua Q, Gao J, Fan ZX. Clinical implications of elevated serum interleukin-6, soluble CD40 ligand, metalloproteinase-9, and tissue inhibitor of metalloproteinase-1 in patients with acute ST-segment elevation myocardial infarction. *Clin Cardiol* 2008, 31(9):413-8.

94. Tan J, Hua Q. Correlations between serum inflammation factors and left ventricular remodeling in acute ST segment elevation myocardial infarction. *Yonsei Med J* 2012, 53(3):501-795. Tátrai E, Ifj Hartyánszky I, Lászik A, Hubay M, Acsády G, Sótonyi P. Molecular biological virus identification in dilated cardiomyopathy. *Orv Hetil* 2007, 148(48):2275-8.

96. Tátrai E, Hartyánszky I Jr, Lászik A, Acsády G, Sótonyi P, Hubay M. The role of viral infections in the development of dilated cardiomyopathy. *Pathol Oncol Res* 2011, 17(2):229-35.

97. Terry DF, Evans JC, Pencina MJ, Murabito JM, Vasan RS, Wolf PA, Kelly-Hayes M, Levy D, D'Agostino RB Sr, Benjamin EJ. Characteristics of Framingham offspring participants with long-lived parents. *Arch Intern Med* 2007, 167:438–44.

98. Thompson PD, Buchner D, Pina IL, Balady GJ, Williams MA, Marcus BH, Berra K, Blair SN, Costa F, Franklin B, Fletcher GF, Gordon NF, Pate RR, Rodriguez BL, Yancey AK, Wenger NK; American Heart Association Council on Clinical Cardiology Subcommittee on Exercise, Rehabilitation, and Prevention; American Heart Association Council on Nutrition, Physical Activity and Metabolism Subcommittee on Physical Activity. Exercise and physical activity in the prevention and treatment of atherosclerotic cardiovascular disease: a statement from the Council on Clinical Cardiology (Subcommittee on Exercise, Rehabilitation, and Prevention). *Circulation* 2003, 107(24):3109-16.

99. Tousoulis D, Antoniadis C, Koumallos N, Stefanadis C. Pro-inflammatory cytokines in acute coronary syndromes: from bench to bedside. *Cytokine Growth Factor Rev* 2006, 17(4):225-33.

100. Tremolada S, Delbue S, Ferrareso M, Carloni C, Elia F, Larocca S, Bortolani E, Ferrante P. Search for genomic sequences of microbial agents in atherosclerotic plaques. *Int J Immunopathol Pharmacol* 2011, 24(1):243-6.

101. Utermann G, Hardewig A, Zimmer F. Apolipoprotein E phenotypes in patients with myocardial infarction. *Hum Genet* 1984, 65(3):237-41.
102. Vaccarino L, Vitale S, Caruso M, Palmeri M, Scola L, Bova M, Caruso C, Massenti MF, Vitale F, Novo S, Lio D, Forte GI. Myocardial infarction marker levels are influenced by prothrombin and tumor necrosis factor- α gene polymorphisms in young patients. *Cytokine* 2013, 61(1):218-22.
103. Van der Linde K, Boor PP, Sandkuijl LA, Meijssen MA, Savelkoul HF, Wilson JH, De Rooij FW. A Gly15Arg mutation in the interleukin-10 gene reduces secretion of interleukin-10 in Crohn disease. *Scand J Gastroenterology* 2003, 38:611-7.
104. Vasan RS, Larson MG, Leip EP, Evans JC, O'Donnell CJ, Kannel WB, Levy D Impact of high-normal blood pressure on the risk of cardiovascular disease. *N Engl J Med* 2001, 345(18):1291-7.
105. Wang TJ, Gona P, Larson MG, Tofler GH, Levy D, Newton-Cheh C, Jacques PF, Rifai N, Selhub J, Robins SJ, Benjamin EJ, D'Agostino RB, Vasan RS. Multiple biomarkers for the prediction of first major cardiovascular events and death. *N Engl J Med* 2006, 355(25):2631-9.
106. Wilhelmsen L, Svärdsudd K, Korsan-Bengtson K, Larsson B, Welin L, Tibblin G. Fibrinogen as a risk factor for stroke and myocardial infarction. *N Engl J Med* 1984, 311(8):501-5.
107. World Health Organization. The top 10 causes of death, factsheet No. 310. <http://who.int/mediacentre/factsheets/fs310/en/index.html>, Retrieved 14 August 2013
108. Xenaki E, Hassoulas J, Apostolakis S, Sourvinos G, Spandidos DA. Detection of cytomegalovirus in atherosclerotic plaques and nonatherosclerotic arteries. *Angiology* 2009, 60(4):504-8.
109. Yao K, Crawford JR, Komaroff AL, Ablashi DV, Jacobson S. Review part 2: human herpesvirus-6 in central nervous system diseases. *J Med Virol* 201, 82: 1669-78.
110. Zhang X, Zhang B, Zhang M, Han Y, Zhao Y, Meng Z, Li X, Kang J, Yan C. Interleukin-8 gene polymorphism is associated with acute coronary syndrome in the Chinese Han population.

Cytokine 2011, 56(2):188-91.

111. Zhou RH, Shi Q, Gao HQ, Shen BJ. Changes in serum interleukin-8 and interleukin-12 levels in patients with ischemic heart disease in a Chinese population. *J Atheroscler Thromb* 2001, 8(1):30-2.

112. Zureik M, Czernichow S, Courbon D, Blacher J, Ducimetière P, Hercberg S, Safar ME, Galan P. Parental longevity, carotid atherosclerosis, and aortic arterial stiffness in adult offspring. *Stroke* 2006, 37(11):2702-7.